



Zooplankton diel vertical migration during Antarctic summer

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ABSTRACT

Zooplankton diel vertical migration (DVM) during summer in the polar oceans is presumed to be dampened due to near continuous daylight. We analyzed zooplankton diel vertical distribution patterns in a wide range of taxa along the Western Antarctic Peninsula (WAP) to assess if DVM occurs, and if so, what environmental controls modulate DVM in the austral summer. Zooplankton were collected during January and February in paired day-night, depth-stratified tows through the mesopelagic zone along the WAP from 2009–2017, as well as in day and night epipelagic net tows from 1993–2017. The copepod *Metridia gerlachei*, salp *Salpa thompsoni*, pteropod *Limacina helicina antarctica*, and ostracods consistently conducted DVM between the mesopelagic and epipelagic zones. Migration distance for *M. gerlachei* and ostracods decreased as photoperiod increased from 17 to 22 h daylight. The copepods *Calanoides acutus* and *Rhincalanus gigas*, as well as euphausiids *Thysanoessa macrura* and *Euphausia crystallorophias*, conducted shallow (mostly within the epipelagic zone) DVMs into the upper 50 m at night. *Rhincalanus gigas*, *T. macrura*, and *L. h. antarctica* DVM behavior was modulated by chlorophyll *a* concentration, mixed layer depth, and depth of the subsurface chlorophyll *a* maximum, respectively. Carnivorous and detritivorous taxa – including the calanoid copepod *Paraeuchaeta antarctica*, ostracods, chaetognaths, and *Tomopteris* spp. polychaetes – as well as seasonally migrating copepods, were most abundant in the mesopelagic zone regardless of the diel cycle. *Paraeuchaeta antarctica* underwent reverse DVM within the top 100 m. The impacts of Antarctic zooplankton summer DVM and the resident mesopelagic assemblage on carbon export should be better quantified.

1. Introduction

Many zooplankton and fishes throughout the world's oceans undergo diel vertical migration (DVM), feeding in productive surface waters at night and seeking refuge from visual predators at mesopelagic depths during the daytime (Hays, 2003). A global estimate suggests ~50% of sound-scattering mesopelagic biomass performs DVM (Klevjer et al., 2016). Diverse zooplankton taxa independently evolved DVM behavior, as it optimizes the adaptive balance between feeding and predator evasion (Zaret and Suffern, 1976; Stich and Lampert, 1981; Gliwicz, 1986; Hays, 2003). While predator avoidance is the accepted evolutionary driver for DVM, shifts in downwelling irradiance at sunrise and sunset are the dominant proximate cues for this behavior (Ringelberg and Van Gool, 2003; Cohen and Forward, 2009). Thus, it was assumed that DVM is restricted in polar regions and may cease altogether in mid-winter and summer during 24-h darkness and light, respectively

(Blachowiak-Samolyk et al., 2006).

DVM studies in the polar oceans show seasonal variability in behavior, with DVM magnitude changing in relation to photoperiod. During moored Acoustic Doppler Current Profiler deployments in the Ross, Lazarev, and Weddell Seas, DVM continued through Antarctic winter but ceased during the period of extended daylight from November to February (Cisewski et al., 2010; Cisewski and Strass, 2016; Picco et al., 2017). Persistent winter DVM occurs as far north as 77°N (Hobbs et al., 2018), although zooplankton DVM is restricted to small-scale (6 to 8 m) migrations within the upper 30 m during Arctic winter (Ludvigsen et al., 2018). Therefore, the apparent pause of DVM during Antarctic summer may in fact be due to seasonal changes in DVM amplitude as some species undertake shallower migrations during summer that go undetected by conventional sampling (Flores et al., 2014; Daase et al., 2016). Asynchronous migrations throughout the diel cycle may also explain why acoustic records do not detect summer DVM

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in the Southern Ocean (Cottier et al., 2006).

In addition to photoperiod, other environmental conditions are likely to influence the amplitude of zooplankton DVM during polar summer. Phytoplankton blooms may halt DVM as zooplankton remain in surface waters to feed (Cisewski et al., 2010; Cisewski and Strass, 2016). The depth of the subsurface chlorophyll *a* maximum can also influence zooplankton DVM, as observed with Arctic copepods (La et al., 2015a). Similarly, vertically migrating Arctic zooplankton concentrate just below the mixed layer (Berge et al., 2014). In the southern California Current, increased light attenuation results in a decreased amplitude of copepod DVM (Ohman and Romagnan, 2016). The interaction between light conditions and phytoplankton distribution is thus likely to be a key driver of Antarctic zooplankton vertical distribution.

Few studies address polar mesopelagic zooplankton composition and taxon-specific variability in DVM behavior. Arctic zooplankton demonstrate asynchronous DVM patterns, which are explained by variation in feeding ecology, predation risk, and seasonal migration behavior (Fortier et al., 2001; Cottier et al., 2006; Falk-Petersen et al., 2008). Prior studies assessing zooplankton vertical distribution along the Western Antarctic Peninsula (WAP) have either focused on specific taxonomic groups (e.g., Nordhausen, 1994a; Lopez and Huntley, 1995) or lacked the comparable day and night sampling necessary to assess

DVM behavior (Marrari et al., 2011). Additionally, there have not been any comprehensive studies resolving polar DVM variability over inter-annual timescales.

We analyzed zooplankton diel vertical distribution patterns along the WAP during mid-summer as part of the Palmer Antarctica Long-Term Ecological Research (PAL LTER) program. Zooplankton were sampled at discrete depth intervals through the epi- and mesopelagic zones in paired day and night net tows using a MOCNESS (2009–2017) to assess taxon-specific zooplankton vertical distribution. Day and night epipelagic net tows (1993–2017) throughout the PAL LTER sampling region provided additional information on DVM behavior. We examined environmental controls (e.g., photoperiod, mixed layer depth) on DVM amplitude in taxa showing clear DVM. Our results show diverse zooplankton DVM modes, depth distributions, and responses to phytoplankton biomass and vertical distribution, all of which can affect zooplankton-mediated carbon export during Antarctic summer.

2. Materials and methods

2.1. Study region

The PAL LTER study region ranges from Anvers Island (64.77°S,

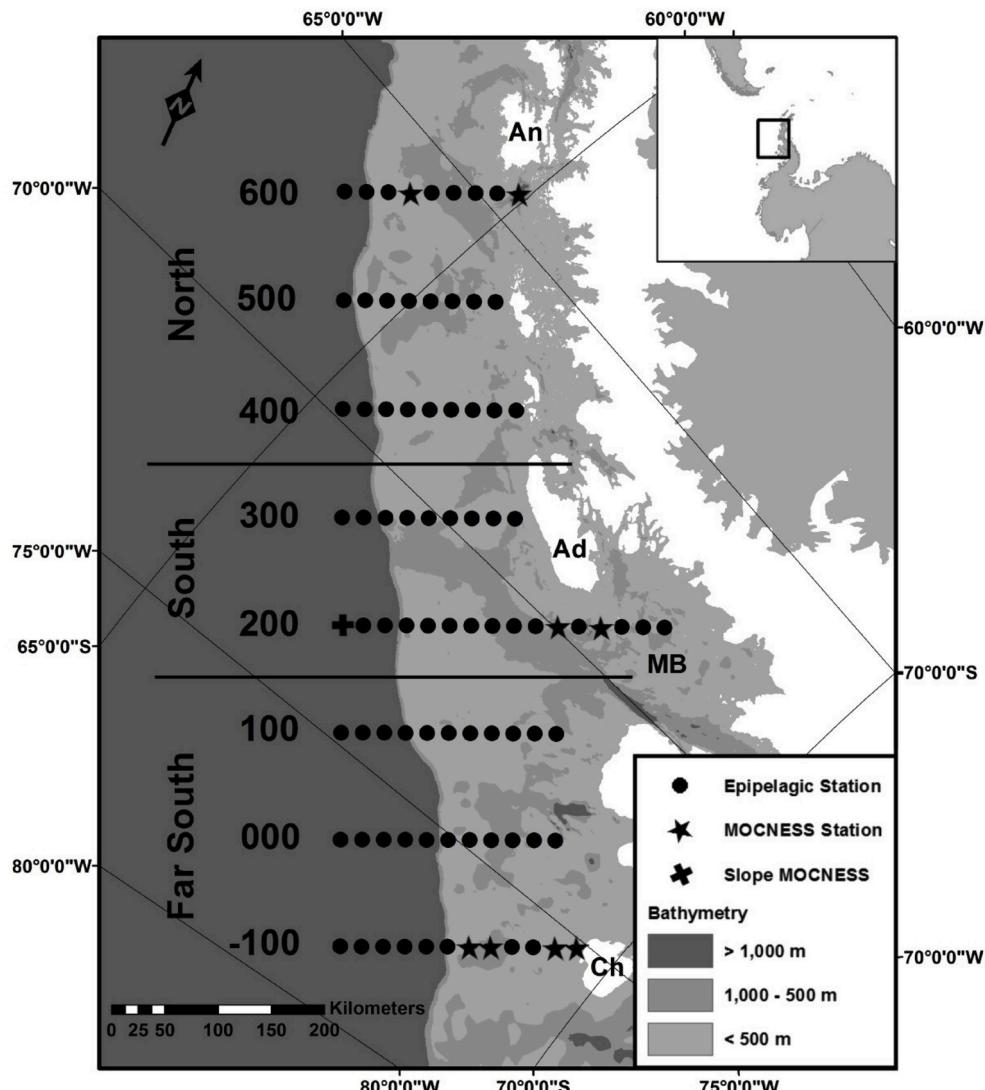


Fig. 1. PAL LTER study area along the Western Antarctic Peninsula. ‘North,’ ‘South,’ and ‘Far South’ regions are indicated. Circles indicate epipelagic sampling stations (1993–2017). Stars indicate paired day-night MOCNESS sampling locations on the shelf (2009–2017). Cross indicates paired day-night MOCNESS sampling station on the slope (2017). Shading indicates bathymetry. An: Anvers Island; Ad: Adelaide Island; MB: Marguerite Bay; Ch: Charcot Island.

64.05°W) in the north to Charcot Island (69.45°S, 75.15°W) in the south, extending from the WAP coast to the continental slope (Ducklow et al., 2012) (Fig. 1). The PAL LTER research grid is composed of sampling lines running perpendicular to the Peninsula every 100 km, and standard grid stations within each line are separated by 20 km (Waters and Smith, 1992). From 1993–2008, the sampling plan included all stations on grid lines 600 to 200. In more recent years, the study area has expanded to include lines 100, 000, and –100, with sampling resolution reduced to three stations per line. As in previous studies, three latitudinal sub-regions were designated to represent hydrographic, sea ice, and ecological gradients (Martinson et al., 2008; Stammerjohn et al., 2008; Steinberg et al., 2015) (Fig. 1), with regional boundaries along sampling grid lines as follows: ‘North’ (lines 600 to 400), ‘South’: (lines 300 and 200), and ‘Far South’ (lines 100 to –100).

2.2. Zooplankton collection

Zooplankton were sampled during austral summer (02 January to 13 February) on annual PAL LTER research cruises aboard the MV *Polar Duke* (1993–1997) and ARSV *Laurence M. Gould* (1998–2017). Zooplankton sampling was conducted using two types of gear as described below.

2.2.1. Multiple discrete depth sampling through the epi- and mesopelagic zones

A 1.4-m² frame, 500-μm mesh Multiple Opening/Closing Net and Environmental Sensing System (MOCNESS) (Wiebe et al., 1985) was used to collect meso- and macrozooplankton in discrete depth intervals from 2009–2017. Each year, paired day (10:07–15:15 local start time) and night (23:03–01:46) MOCNESS tows were carried out in coastal or shelf waters in the ‘North’, and in most years also the ‘South’ and ‘Far South’ (Fig. 1). Sampling time and location were used to calculate solar elevation at the start of tows (Meeus, 1998); daytime solar elevation was >35.51° and nighttime solar elevation < 0.56°. The MOCNESS was towed obliquely at a speed of 2–2.5 knots, with a typical tow duration of 2.25–3 h. On average, 790 m³ (range 161–1900 m³) of water was filtered within a single depth interval as measured by a flow meter mounted on the system.

Eight discrete depth intervals were sampled during the upcast as follows: 500–400, 400–300, 300–250, 250–200, 200–150, 150–100, 100–50, and 50–0 m. Occasionally, the deeper intervals were not sampled when towing in waters shallower than the deep target depths. On four occasions, two depth intervals were combined due to net sampling errors. In these cases, the taxon density in each interval was assumed to equal the density calculated for the combined interval. Sea ice conditions occasionally prevented MOCNESS sampling in the South and Far South regions, therefore, the sample size $n = 7$ for paired day/night tows in the North, while $n = 6$ for the South, and $n = 4$ for the Far South from 2009–2015. Data from additional paired MOCNESS tows in 2016 and 2017 are presented for euphausiids and salps only (for these taxa, $n = 10$ for the North, $n = 8$ for the South, and remains $n = 4$ for the Far South).

A pair of day (09:54 local start time) and night (22:03) tows was also carried out over the continental slope in 2017 (Fig. 1). In this case, eight discrete depth intervals were sampled during the upcast as follows: 1000–750, 750–500, 500–400, 400–300, 300–200, 200–100, 100–50, and 50–0 m (100–0 m for day, due to a net sampling error). Salp data from this slope sample are presented independent of the coastal and shelf data.

2.2.2. Epipelagic sampling

Macrozooplankton were also collected from 1993–2017 throughout the PAL LTER study area using a 2 × 2 m square, 700-μm mesh Metro net towed obliquely to 120 m (Ross et al., 2008; Steinberg et al., 2015) (Fig. 1). The net depth and tow profile were monitored with a depth sensor linked to the conducting hydro wire. Average volume filtered was

9023 m³ (range 1715–71929 m³), calculated using a General Oceanics flow meter suspended in the net opening. Ship speed was 2–2.5 knots while towing, and typical tow duration was 30–35 min.

Epipelagic samples were designated night tows when the sun was below the horizon, accounting for atmospheric refraction (calculated solar elevation $\leq -0.833^\circ$ at the start of the tow) (Atkinson et al., 2008; Steinberg et al., 2015). Sample size varied by taxon (day $n = 966$ –1071 and night $n = 181$ –198), as not all taxa were identified during shipboard processing throughout the time series.

2.3. Taxonomic composition

2.3.1. Discrete depth samples

All euphausiids and salps collected in MOCNESS tows were identified and quantified at sea. Whole samples were then preserved in sodium borate-buffered 4% formaldehyde and shipped to the Virginia Institute of Marine Science (Gloucester Point, VA, USA) for further taxonomic analysis to quantify all non-euphausiid or salp taxa. Samples were size-fractionated using a 5-mm mesh, with all individuals in this larger size fraction identified and counted. This size-fractionation step removed large, abundant taxa (i.e., salps and euphausiids) from the microscopic analysis. At least 1/64 of the < 5-mm size fraction was counted under a stereo dissecting microscope after dividing the sample with a plankton splitter. A minimum of 100 individuals of the most abundant species was enumerated in this smaller size fraction.

Discrete depth analyses focused on abundant taxonomic groups. Five common calanoid copepod species were included: *Metridia gerlachei*, *Calanoides acutus*, *Calanus propinquus*, *Rhincalanus gigas*, and *Paraeuchaeta antarctica*. Identification included adults and conspicuous copepodites. Discrete depth-stratified data were only analyzed for the smaller, but abundant, euphausiids *Thysanoessa macrura* and *Euphausia crystallorophias*, because the larger Antarctic krill *Euphausia superba* was underrepresented due to avoidance of the 1.4-m² MOCNESS (Nordhausen, 1994b). The pelagic tunicate *Salpa thompsoni* was included. The thecosome (shelled) pteropod *Limacina helicina antarctica* was analyzed individually while the gymnosome (shell-less) pteropods *Clione antarctica* and *Spongibranchea australis* were grouped together (as in Thibodeau et al., 2019). *Tomopteris* spp. polychaetes were combined into a single group including *T. carpenteri*. Other groups were analyzed by major taxa, such as ostracods, amphipods (including the hyperiids *Themisto gaudichaudii*, *Cyllopodus lucasi*, *Hyperiella macronyx*, *Hyperoche medusarum*, *Primno macropa*, *Vibiliab stebbingi*, *Scina* spp., and the gammarid *Eusirus* spp.), and chaetognaths (inclusive of large, conspicuous *Pseudosagitta gazellae* and *P. maxima*).

2.3.2. Epipelagic samples

Grid-wide epipelagic tows were sorted at sea as reported in Steinberg et al. (2015). All above taxa were included in the analysis of epipelagic samples except for the calanoid copepods and ostracods. The Antarctic krill *Euphausia superba* was included in analysis of epipelagic samples.

2.4. Vertical structure

Night to day ratios (N:D) were calculated to identify diel changes in surface abundance of each taxon. For paired day and night MOCNESS samples, abundance was integrated to 150 m and to 50 m (individuals m⁻²) when a taxon was present in both the day and night tows (Steinberg et al., 2008). These values are referred to as MOCNESS 150 m N:D and MOCNESS 50 m N:D, respectively. MOCNESS N:D data typically ranged across multiple orders of magnitude and were positively skewed. A relatively few large values were influential on the mean MOCNESS N:D values, typically resulting in large mean values compared to the median (Supplemental Tables 1 and 2). Therefore, the median was used to describe the central tendency of MOCNESS N:D data. Additionally, mean day abundance and mean night abundance were calculated from epipelagic (0–120 m) samples to calculate grid-wide N:D ratios for

1993–2017. These values are referred to as grid-wide 120 m N:D.

The vertical distribution of taxa in the MOCNESS discrete depth samples was quantified using weighted mean depth (WMD). WMD (m) is calculated as follows:

$$WMD = \sum (n_i \times z_i \times d_i) / \sum (n_i \times z_i)$$

where for depth interval i , d_i is the midpoint (m), z_i is the interval thickness (m), and n_i is abundance (no. m^{-3}) (Andersen et al., 2001). WMD was only calculated for tows reaching 500 m. Night WMD was subtracted from day WMD to determine the amplitude of diel migration (Δ WMD, m). Data used in the analyses are available at: <https://pal.lternet.edu/data>.

2.5. Environmental controls

The environmental water column data used in this analysis was collected at sampling stations where paired day-night MOCNESS tows were conducted. Discrete chlorophyll *a* (chl-*a*) measurements were made fluorometrically (Parsons et al., 1984). Primary productivity rates were measured with 24-h incubations of ^{14}C uptake at various light levels (Steemann Nielsen, 1952; Schofield et al., 2018). Both chl-*a* and primary production were depth-integrated to 100 m. The depth of the subsurface chl-*a* maximum (Z_{SCM}) and euphotic zone defined by the 1% isolume ($Z_{1\%}$) were determined with a fluorometer and a photosynthetically active radiation (PAR) sensor, respectively, mounted on the CTD rosette. Mixed layer depth (MLD) was calculated as the depth of maximum buoyancy frequency from the same CTD casts (Carvalho et al., 2017). Photoperiod (hours) was calculated for all day-night MOCNESS tow pairs using latitude and day of year (Kirk, 2011).

2.6. Statistical analyses

Single-factor ANOVA was used to test for differences in Δ WMD and log-adjusted MOCNESS N:D ratios among the North, South, and Far South sub-regions. The significance level (α) was set at 0.05. There was no significant difference among latitudinal sub-regions for any taxa for MOCNESS 150 m N:D or MOCNESS 50 m N:D (ANOVA; $p > 0.06$). All statistical tests were conducted with R version 3.3.2 (R Core Team, 2016).

Differences between day and night surface abundance (0–150 m and 0–50 m) from MOCNESS pairs were tested using the Wilcoxon signed-rank test. This non-parametric test does not require transformation of non-normal data and gave comparable results to the paired *t*-test using log-transformed data (Supplemental Tables 1 and 2). Differences between unpaired day and night grid-wide epipelagic abundance (0–120 m) were tested using the Wilcoxon rank-sum test.

Multiple linear regression was used to identify environmental controls on Δ WMD for taxa that made DVMs from the mesopelagic zone into the epipelagic zone. Only *M. gerlachei* and ostracods were included in this analysis; Δ WMD was not a sensitive metric for *L. h. antarctica*, because it was concentrated in the epipelagic zone, and *Salpa thompsoni* was excluded due to an insufficient sample size. Δ WMD did not differ among latitudinal sub-regions for *M. gerlachei* or ostracods (ANOVA; $p > 0.96$). Therefore, data for the analysis were combined across the entire sampling region.

MOCNESS 50 m N:D was a sensitive metric for taxa that were concentrated in the epipelagic zone or conducted DVM within the epipelagic zone. These taxa included *L. h. antarctica*, *C. acutus*, *R. gigas*, *T. macrura*, and *E. crystallorophias*. Generalized linear models with a gamma distribution and log link function were used to identify environmental controls on MOCNESS 50 m N:D. The gamma distribution is appropriate for ratios, because it is constrained to positive, continuous values. The log link function also ensures positive fitted values. Significant model fits were not achieved for the shallow migrants *C. acutus* and *E. crystallorophias*.

A suite of nine models was fitted for each individual taxon included in Δ WMD analysis (multiple linear regression) and in MOCNESS 50 m N:D analysis (generalized linear model with gamma distribution and log link function). The water column properties investigated in this study were correlated with one another and therefore were not included in the same models to avoid problematic collinearity. For example, as MLD deepened so did Z_{SCM} (Pearson's $r = 0.69$; $p = 0.0004$). $Z_{1\%}$ deepened as depth-integrated chl-*a* decreased (Pearson's $r = -0.60$; $p = 0.004$) and as Z_{SCM} deepened (Pearson's $r = 0.44$; $p = 0.044$). None of the water column properties were correlated with photoperiod. Therefore, the nine models included each explanatory variable individually (i.e., photoperiod, chl-*a*, Z_{SCM} , $Z_{1\%}$, and MLD) as well as photoperiod paired with each of the water column properties. Model selection statistics are presented in Supplemental Tables 3–7. Final models were selected according to the lowest Akaike Information Criterion value corrected for small sample size (AICc) (Hurvich and Tsai, 1989) using the model.sel function in the MuMin package (Barton, 2016). Presented models satisfied assumptions as verified by plotting residuals versus fitted values and explanatory variables.

3. Results

3.1. Environmental conditions

Mean photoperiod during MOCNESS sampling was 20 h 11 min (range: 17 h 41 min to 21 h 47 min), and mean $Z_{1\%}$ was 47 m (range: 16–81 m). Mean depth-integrated chl-*a* was 126 mg m^{-2} (range: 13–517 mg m^{-2}), and mean depth-integrated primary production was 2489 mg C $m^{-2} d^{-1}$ (range: 605–5354 mg C $m^{-2} d^{-1}$). Mean MLD was 28 m (range: 5–79 m), and mean Z_{SCM} was 20 m (range: 4–60 m).

3.2. Diel vertical depth distributions by taxon

3.2.1. Calanoid copepods

The calanoid copepod *Metridia gerlachei* was the most abundant taxon in MOCNESS tows and a strong diel vertical migrator (Fig. 2), with a median MOCNESS 150 m N:D of 8.0 (Wilcoxon signed-rank test $p = 0.002$) (Table 1). Much of the *M. gerlachei* population did not migrate and resided between 300–500 m, particularly in the North (Fig. 2a). The *M. gerlachei* depth distribution was more even in the South and Far South (Fig. 2b–c). *Calanoides acutus* was the second-most abundant calanoid and although its abundance from 0–150 m did not differ significantly between day and night (Wilcoxon signed-rank test $p = 0.64$) (Table 1), it was more abundant during night tows from 0–50 m with a median MOCNESS 50 m N:D of 2.3 (Fig. 3d; Table 2) (Wilcoxon signed-rank test $p = 0.001$). Like *M. gerlachei*, *C. acutus* vertical distribution also varied with latitudinal sub-region. *Calanoides acutus* was distributed relatively evenly with depth in the North and South (Fig. 3a–b) but was concentrated between 250–400 m in the Far South (Fig. 3c), where it was also an order of magnitude more abundant at this depth zone compared to the other sub-regions.

In contrast to *M. gerlachei* and *C. acutus*, *Calanus propinquus*, *Rhincalanus gigas*, and *Paraeuchaeta antarctica* were an order of magnitude less abundant and did not vary appreciably with latitudinal sub-region. *Calanus propinquus* was most abundant in the surface 50 m (Fig. 4a) unlike other calanoid copepods, which had peak abundances in the mesopelagic zone (day and night). Epipelagic *C. propinquus* abundance did not differ between day and night (Tables 1 and 2). *Rhincalanus gigas* was most abundant from 250–300 m during the day and from 200–250 m at night (Fig. 4b), and abundance in the surface 50 m was significantly greater at night than during the day (Wilcoxon signed-rank test $p = 0.004$). Median *R. gigas* MOCNESS 50 m N:D was 36.6 (Table 2). *Paraeuchaeta antarctica* was most abundant from 300–500 m and mostly remained resident in the mesopelagic zone during day and night (Fig. 4c). Although scarce in the epipelagic zone, *P. antarctica* was significantly more abundant from 0–50 m during the day (Wilcoxon

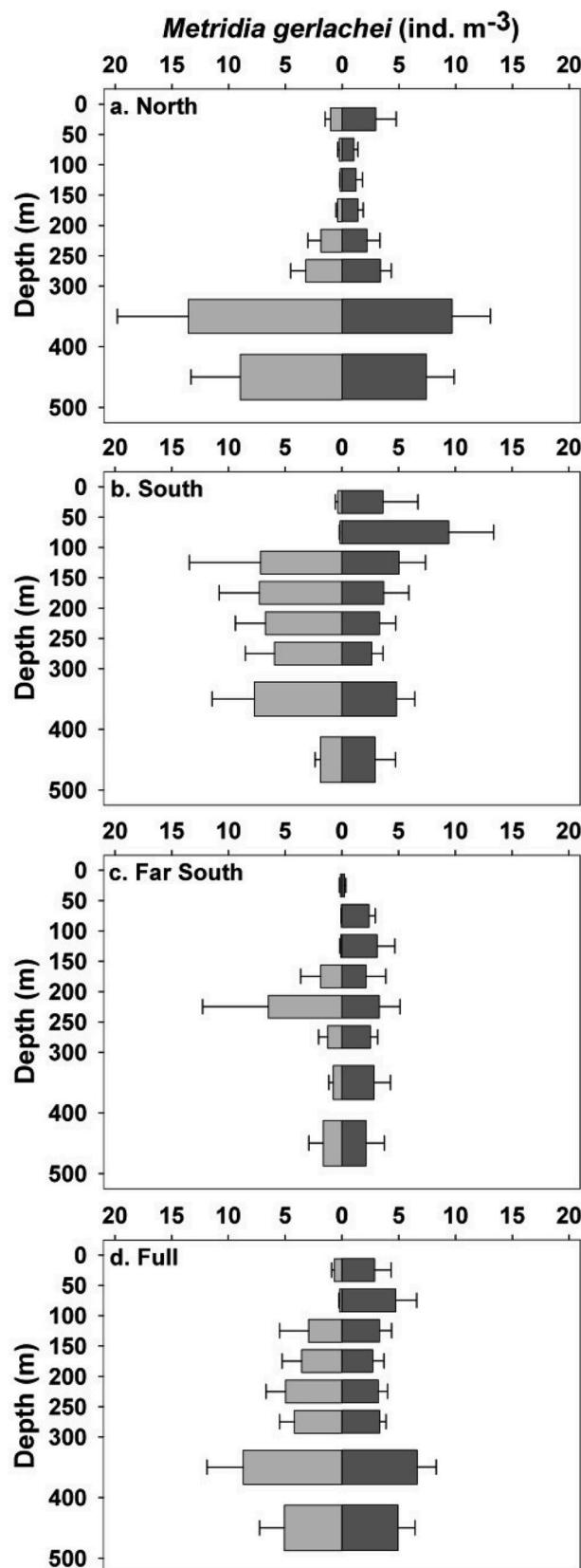


Fig. 2. Mean day (light gray, left) and night (dark gray, right) abundance of the calanoid copepod *Metridia gerlachei* in the North (a), South (b), Far South (c) sub-regions, and full shelf sampling region (d) at discrete depth intervals from 0–500 m. Error bars indicate one standard error. North $n = 5\text{--}7$; South $n = 5\text{--}6$; Far South $n = 2\text{--}4$; Full $n = 12\text{--}17$.

signed-rank test $p = 0.012$) (Table 2), which suggests this species conducted reverse DVM.

3.2.2. Euphausiids

The abundant krill species *Thysanoessa macrura* was concentrated in the epipelagic zone during day and night, but ascended at night, especially into the upper 50 m (Fig. 5a). Some degree of DVM by *T. macrura* was supported by all metrics tested. This species was significantly more abundant in 0–50 m depths at night compared to day (Wilcoxon signed-rank test $p = 0.006$) (Table 2), and *T. macrura* median MOCNESS 50 m N:D was 11.7. *Thysanoessa macrura* was also more abundant at night vs. day in the upper 0–150 m (MOCNESS tows; Wilcoxon signed-rank test $p = 0.003$; median MOCNESS 150 m N:D = 1.4) (Table 1) and from 0–120 m during nighttime grid-wide epipelagic tows (Wilcoxon rank-sum test $p = 0.0003$; grid-wide 120 m N:D = 1.6) (Table 3). *Euphausia crystallorophias* was less abundant than *T. macrura*, but similarly was concentrated in the upper 100 m (Fig. 5b) and migrated into the top 50 m at night, as indicated by higher abundance from 0–50 m at night than day (Wilcoxon signed-rank test $p = 0.047$) and a median MOCNESS 50 m N:D of 8.8 (Table 2). *Euphausia superba* remained in the epipelagic zone through the diel cycle with a grid-wide 120 m N:D of 0.93 (Table 3).

3.2.3. Other crustaceans

Ostracods migrated nightly into the upper 200 m (Fig. 6a), with significantly higher abundance from 0–150 m at night vs. day (Wilcoxon signed-rank test $p = 0.0003$) and a median MOCNESS 150 m N:D of 2.9 (Table 1). Mean ostracod abundance peaked in the 200–250 m layer, where they were about 50% more abundant during day than night (Fig. 6a). Most of the ostracod community did not migrate and resided between 200–500 m throughout the diel cycle. Amphipods were an order of magnitude less abundant than ostracods, with two distinct abundance peaks in the mesopelagic zone during day, and highest abundance from 100–200 m at night (Fig. 6b). Amphipods were significantly more abundant in nighttime epipelagic tows and had a grid-wide 120 m N:D of 2.1 (Wilcoxon rank-sum test $p = 2.6 \times 10^{-12}$) (Table 3).

3.2.4. Salps

Salpa thompsoni was a strong diel vertical migrator. Mean abundance over the continental shelf was highest from 200–400 m during the day, and from 0–200 m at night (Fig. 7a). *Salpa thompsoni* median MOCNESS 150 m N:D was 9.0, although salps were only present in three day-night MOCNESS tow pairs along the continental shelf (Table 1). Salps were also significantly more abundant at night in epipelagic tows, with a grid-wide 120 m N:D of 2.6 (Wilcoxon rank-sum test $p = 7.7 \times 10^{-17}$) (Table 3). Over the continental slope, *S. thompsoni* migrated mostly from daytime residence depths in the 200–300 m layer into the surface 100 m at night (Fig. 7b), with MOCNESS 100 m N:D = 94.6 and MOCNESS 200 m N:D = 6.3. Salps were relatively scarce below 300 m, although a small, deep peak occurred from 750–1000 m on the slope.

3.2.5. Pteropods

The thecosome (shelled) pteropod *Limacina helicina antarctica* was concentrated in surface waters but also migrated from 150–250 m during the day into the upper epipelagic zone at night (Fig. 8a). This result is supported by their higher abundance in the upper 150 m at night from MOCNESS tows (Wilcoxon signed-rank test $p = 0.021$) (Table 1) and from 0–120 m at night in epipelagic tows (Wilcoxon rank-sum test $p = 0.004$) (Table 3). Median *L. h. antarctica* MOCNESS 150 m N:D was 1.7 (Table 1), and grid-wide 120 m N:D was 1.4 (Table 3). Gymnosome (shell-less) pteropods were less abundant than *L. h. antarctica* in the epipelagic zone and were distributed relatively evenly with depth, with highest mean gymnosome abundance from 0–50 m during the day (Fig. 8b). However, DVM by gymnosomes is indicated grid-wide, with significantly higher abundance at night in the epipelagic zone and a grid-wide 120 m N:D of 1.4 (Wilcoxon rank-sum test $p = 0.0002$)

Table 1

Diel, depth-integrated zooplankton abundance (0–150 m) from MOCNESS tows along the WAP continental shelf. For euphausiids and *Salpa thompsoni*, Day and Night $n = 22$ (samples from 2009–2017). For all other taxa, Day and Night $n = 17$ (samples from 2009–2015). Night:Day n varies because taxa were not always present in both day and night paired tows. p -values are for the Wilcoxon signed-rank test comparing paired day-night abundance values when a taxon was present in both tows.

Taxon	Day (individuals m^{-2})		Night (individuals m^{-2})		Night:Day			p	n
	Median	Range	Median	Range	Median	25%–75% Quantiles			
Calanoid copepods									
<i>Metridia gerlachei</i>	32.7	0.1–1975.0	249.5	3.1–2860.8	8.0	6.2–32.0		0.002	17
<i>Calanoides acutus</i>	37.1	4.2–230.6	51.6	7.3–269.1	1.0	0.8–1.5		0.64	17
<i>Calanus propinquus</i>	4.8	0.0–165.4	7.3	1.0–75.7	1.7	0.4–4.4		0.67	16
<i>Rhincalanus gigas</i>	4.2	0.3–20.3	3.9	0.0–14.8	1.2	0.5–2.3		0.82	16
<i>Paraeuchaeta antarctica</i>	0.9	0.0–4.5	1.4	0.0–7.1	1.1	0.6–2.3		0.68	12
Euphausiids									
<i>Thysanoessa macrura</i>	5.0	0.2–60.4	9.9	0.1–74.9	1.4	1.1–2.5		0.003	22
<i>Euphausia crystallorophias</i>	0.2	0.0–18.3	0.1	0.0–61.6	0.7	0.5–5.6		0.58	11
Other crustaceans									
Ostracoda	3.8	0.0–38.2	20.1	1.2–219.1	2.9	1.7–7.4		0.0003	16
Amphipoda	1.9	0.0–8.5	1.3	0.0–16.4	0.9	0.6–2.1		0.56	15
Gelatinous zooplankton									
<i>Salpa thompsoni</i> ^a	0.0	0.0–19.4	0.0	0.0–174.4	9.0	4.5–9.0		0.75	3
<i>Limacina helicina antarctica</i>	1.2	0.0–53.2	3.4	0.0–56.1	1.7	1.2–3.6		0.021	13
Gymnosomata	0.5	0.0–25.3	0.6	0.0–3.3	0.8	0.5–1.3		0.34	12
Chaetognatha	24.0	0.7–118.6	21.8	1.7–84.1	1.0	0.6–1.6		0.75	17
<i>Tomopteris</i> spp.	0.04	0.0–1.1	0.4	0.0–1.6	1.2	1.1–6.7		0.30	9

^a See Fig. 7b for *Salpa thompsoni* data from the continental slope.

(Table 3).

3.2.6. Gelatinous carnivores

Chaetognaths and *Tomopteris* spp. polychaetes were mostly resident in the mesopelagic zone and relatively scarce from 0–100 m (Fig. 9). Mean abundance of both taxa was highest from 200–250 m during the day and from 150–200 m at night (*Tomopteris* spp. also had a second night peak at 300–400 m) (Fig. 9). Median chaetognath MOCNESS 150 m N:D was 1.0 (Table 1) and although grid-wide 120 m N:D was 0.6, epipelagic abundance did not differ between day and night (Tables 1–3), suggesting chaetognaths did not undergo DVM. However, *Tomopteris* spp. polychaetes did appear to undergo DVM as they were significantly more abundant during night epipelagic tows and had a grid-wide 120 m N:D of 1.9 (Wilcoxon rank-sum test $p = 0.0002$) (Table 3).

3.3. Environmental controls on DVM

For strong migrators traveling between the mesopelagic and epipelagic zones, migration distance (i.e., Δ WMD) was sensitive to photoperiod and vertical water column structure. For the copepod *M. gerlachei*, photoperiod and Z_{SCM} best explained Δ WMD (Table 4), with *M. gerlachei* making shorter vertical migrations as photoperiod grew longer and when Z_{SCM} was shallower (Fig. 10a). Similarly, photoperiod and MLD best explained ostracod Δ WMD (Table 4). Ostracods made shorter DVMs as photoperiod grew longer and when MLD was deeper (Fig. 10b; note—an outlier that was excluded prior to model selection for ostracods is included for visualization in this figure).

For taxa making shorter-distance DVMs mostly within the epipelagic zone, the magnitude of DVM into the surface layer (i.e., MOCNESS 50 m N:D) was best explained by phytoplankton abundance and distribution. The final models each included a different, single explanatory variable for the copepod *R. gigas*, euphausiid *T. macrura*, and pteropod *L. h. antarctica*. *Rhincalanus gigas* DVM into the surface 50 m decreased as depth-integrated chl-a increased (Fig. 11a; Table 5). *Thysanoessa macrura* DVM decreased when MLD was deeper (Fig. 11b), and similarly *L. h. antarctica* DVM decreased when Z_{SCM} was deeper (Fig. 11c – a finding robust to the inclusion of an outlier value – see inset) (Table 5).

4. Discussion

4.1. Zooplankton DVM modes

4.1.1. DVM between epipelagic and mesopelagic zones

Four taxa performed consistent DVM between the mesopelagic zone during day and epipelagic zone at night. The copepod *Metridia gerlachei* migrated into the upper 100 m at night, consistent with results from prior studies in the northern WAP (Hopkins, 1985; Lopez and Huntley, 1995; King and LaCasella, 2003). Similarly, a portion of the ostracod community made relatively extensive (~100 m) DVMs resulting in a 21% decrease in abundance from 200–300 m and a 3.5-fold increase in abundance from 0–200 m at night. Ostracod DVM is well-documented in the Atlantic and Pacific Oceans (Angel, 1979; Steinberg et al., 2008). Population-wide DVM by *Salpa thompsoni* from 300 m into surface waters supports previous observations throughout the Southern Ocean during summer (Piatkowski, 1985; Casareto and Nemoto, 1986; Perissinotto and Pakhomov, 1998; Pakhomov et al., 2011). Although rarely encountered deeper than 300 m during summer, the pteropod *Limacina helicina antarctica* underwent DVM between the epipelagic and upper mesopelagic zones. In the Lazarev Sea, *L. h. antarctica* also conducted DVM from November to February (Hunt et al., 2008). Collectively, *M. gerlachei*, ostracods, *S. thompsoni*, and *L. h. antarctica* constitute an assemblage of strong vertical migrators along the WAP during summer.

Evidence for amphipod, gymnosome pteropod, and *Tomopteris* spp. polychaete DVM was less consistent than for the above species, but these taxa were each more abundant at night compared to day in grid-wide epipelagic tows. All amphipod species were grouped together, but DVM is likely species-specific. For example, the hyperiid amphipod *Cyllopus lucasi* was more abundant through the upper 200 m at night during summer, autumn, and winter in the Lazarev Sea where there was no evidence for DVM by the hyperiids *Hyperiella dilatata* and *Primno macropa* (Flores et al., 2014). *Themisto gaudichaudii* (synonym *Paramythisto gaudichaudii*) is abundant along the WAP (Steinberg et al., 2015), and this amphipod made DVMs from ~200 m to the surface 50 m in the Atlantic Ocean (Williams and Robins, 1981). Prior evidence for *Clione antarctica* and *Spongibranchia australis* DVM is inconsistent (Hunt et al., 2008), but our epipelagic day-night abundance data suggest these gymnosome pteropods conduct DVM, likely to feed on their primary prey *L. h. antarctica* (Lalli and Gilmer, 1989; Van der Spoel and Dadon, 1999) in the epipelagic zone at night. At night, *Tomopteris* spp. polychaete abundance decreased 37% from 200–300 m and increased 3-fold

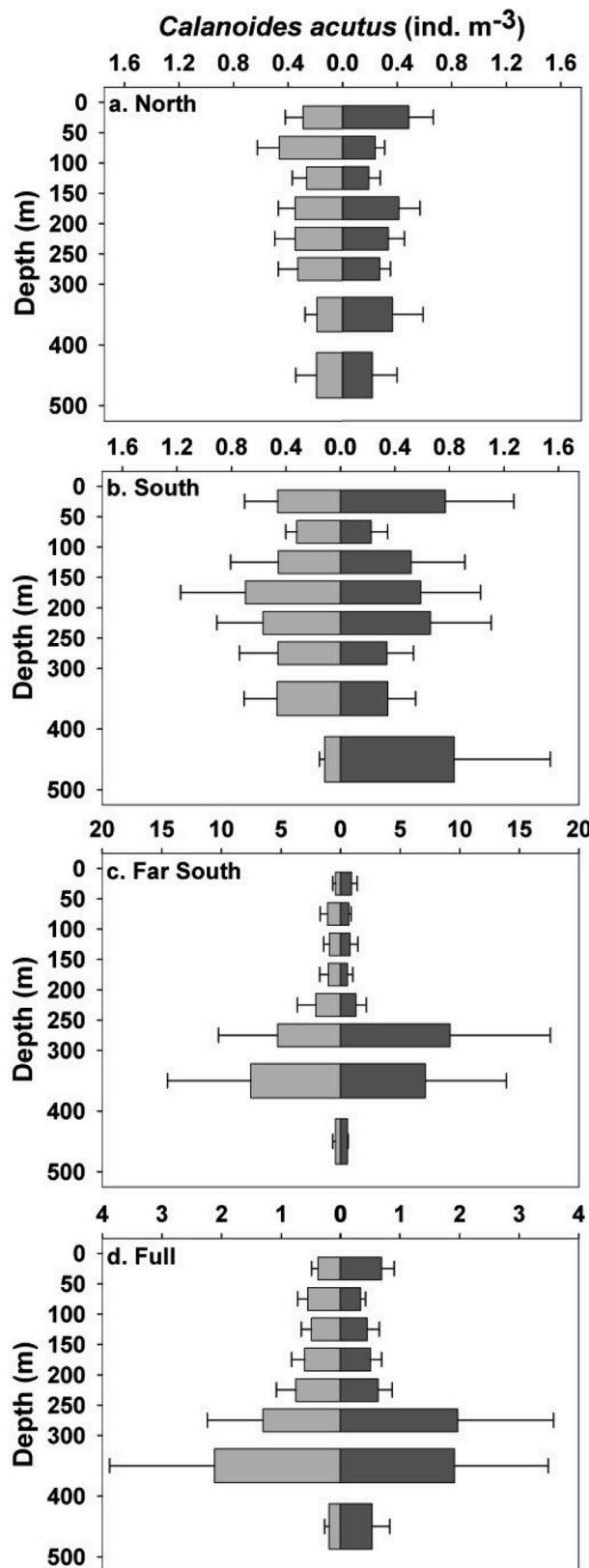


Fig. 3. Mean day (light gray, left) and night (dark gray, right) abundance of the calanoid copepod *Calanoides acutus* in the North (a), South (b), Far South (c) sub-regions, and full shelf sampling region (d) at discrete depth intervals from 0–500 m. Error bars indicate one standard error. North $n = 5–7$; South $n = 5–6$; Far South $n = 2–4$; Full $n = 12–17$. Note different scaling on x-axes.

in the surface 0–200 m. We suggest a portion of the amphipod, gymnosome, and *Tomopteris* spp. assemblage conducted DVM to feed in the upper 200 m while other individuals remained at depth.

4.1.2. DVM within epipelagic zone

The copepods *Calanoides acutus* and *Rhincalanus gigas* made shallow DVMs from the 50–100 m layer into the upper 50 m at night. Shallow DVMs within the upper 70 m for *C. acutus* and upper 90 m for *R. gigas* were also reported in January near South Georgia (Atkinson et al., 1992a, 1992b). A study in the Drake Passage and northern Antarctic Peninsula found no *C. acutus* DVM during December to March (Huntley and Escrictor, 1991), but was limited to vertical resolution of 0–100 and 100–200 m, making it unlikely to detect shallow DVM.

The krill species *Thysanoessa macrura* and *Euphausia crystallorophias* performed shallow DVM. DVM within the epipelagic zone was reported during spring and autumn further north of our study site for *T. macrura* (Loeb and Shulenberger, 1987; Nordhausen, 1994a) and *E. crystallorophias* (using acoustics; Everson, 1987). Summer surveys in the northern WAP (Nordhausen, 1992) and Amundsen Sea (La et al., 2015b) did not detect DVM by *T. macrura* and *E. crystallorophias*, respectively, possibly due to limitations of sampling methods in detecting shallow DVM. Net avoidance by the larger, faster *E. crystallorophias* was not apparent in the northern WAP during winter (Nordhausen, 1994b) but was during autumn (Everson, 1987), which could exaggerate the shallow DVM signal and contribute to higher nighttime abundance depicted in Fig. 5b.

4.1.3. DVM within mesopelagic zone

Chaetognaths and the copepod *R. gigas* both undertook a modest DVM within the mesopelagic zone between 150 and 300 m, possibly indicating predator-prey coupling. The chaetognaths *Eukronia hamata* and *Sagitta gazellae* predominately fed on the copepods *C. acutus*, *C. propinquus*, and *M. gerlachei* in the upper 200 m of the Weddell Sea in autumn (Hopkins and Torres, 1989). Chaetognaths along the WAP mainly remained in deeper layers during summer to feed on abundant copepod prey. Although less numerous, *R. gigas* is a larger (Gleiber, 2014), and perhaps preferable, copepod prey item compared to *M. gerlachei* or *C. acutus*.

4.1.4. Reverse DVM

The copepod *Paraeuchaeta antarctica* made relatively short reverse DVMs, from 0–50 m during the day to 50–100 m at night. The primarily carnivorous *P. antarctica* is the largest copepod in this study (mean adult prosome length 7 mm; Gleiber, 2014), making it particularly vulnerable to visual predators in surface waters (Aksnes and Giske, 1993; Ohman and Romagnan, 2016). Reverse DVM is adaptive for species susceptible to predators that undertake normal DVM (Ohman et al., 1983). The reverse DVM of *P. antarctica* is likely used to avoid vertically migrating visual predators.

4.2. Non-migrating zooplankton

4.2.1. Epipelagic non-migrants

The copepod *Calanus propinquus* was concentrated in the upper 50 m and did not consistently undertake DVM. *Calanus propinquus* feeds omnivorously (Atkinson, 1998; Pasternak and Schnack-Schiel, 2001) and was concentrated in the upper 100 m year-round in the Scotia Sea (Atkinson and Sinclair, 2000). Therefore, it appears *C. propinquus* typically remains resident in surface waters on both seasonal and diel time scales.

The negligible difference between night and day *E. superba* abundance in our 120 m tows was expected since this depth was selected to collect Antarctic krill across its main summer depth range (Ross et al., 1996). Acoustic studies have documented sporadic DVM within the upper 100 m during summer while DVM is more pronounced during spring and autumn (Everson, 1983; Godlewski and Klusek, 1987; Demer

Table 2

Diel, depth-integrated zooplankton abundance (0–50 m) from MOCNESS tows along the WAP continental shelf. For euphausiids and *Salpa thompsoni*, Day and Night $n = 22$ (samples from 2009–2017). For all other taxa, Day and Night $n = 17$ (samples from 2009–2015). Night:Day n varies because taxa were not always present in both day and night paired tows. p -values are for the Wilcoxon signed-rank test comparing paired day-night abundance values when a taxon was present in both tows.

Taxon	Day (individuals m^{-2})		Night (individuals m^{-2})		Night:Day			p	n
	Median	Range	Median	Range	Median	25%–75% Quantiles			
Calanoid copepods									
<i>Metridia gerlachei</i>	4.0	0.0–153.8	7.6	0.0–948.6	1.8	1.4–7.0		0.013	13
<i>Calanoides acutus</i>	9.0	0.0–76.0	13.8	0.0–155.5	2.3	1.6–2.9		0.001	12
<i>Calanus propinquus</i>	0.2	0.0–159.5	4.0	0.0–62.0	2.1	0.9–4.2		0.38	8
<i>Rhincalanus gigas</i>	0.3	0.0–4.9	13.8	0.0–155.5	36.6	18.2–112.4		0.004	9
<i>Paraeuchaeta antarctica</i>	4.0	0.0–62.0	0.1	0.0–1.6	0.2	0.1–0.4		0.012	9
Euphausiids									
<i>Thysanoessa macrura</i>	0.2	0.0–54.8	3.1	0.1–47.1	11.7	3.6–19.4		0.006	16
<i>Euphausia crystallorophias</i>	0.0	0.0–4.7	0.1	0.0–56.3	8.8	4.3–13.1		0.047	7
Other crustaceans									
Ostracoda	1.0	0.0–23.3	0.9	0.0–24.0	1.3	1.0–2.1		0.24	11
Amphipoda	0.3	0.0–8.0	0.3	0.0–3.2	0.7	0.5–2.4		0.85	10
Gelatinous zooplankton									
<i>Salpa thompsoni</i>	0.0	0.0–5.4	0.0	0.0–63.0	1.2	0.6–6.5		0.75	3
<i>Limacina helicina antarctica</i>	0.6	0.0–28.4	0.9	0.0–31.7	2.8	1.7–3.6		0.13	9
Gymnosomata	0.0	0.0–25.0	0.1	0.0–2.5	1.2	0.4–1.3		1.00	6
Chaetognatha	1.9	0.0–14.2	1.6	0.0–10.4	0.9	0.7–1.5		0.68	13
<i>Tomopteris</i> spp.	0.0	0.0–1.1	0.0	0.0–0.8	0.5	NA		NA	1

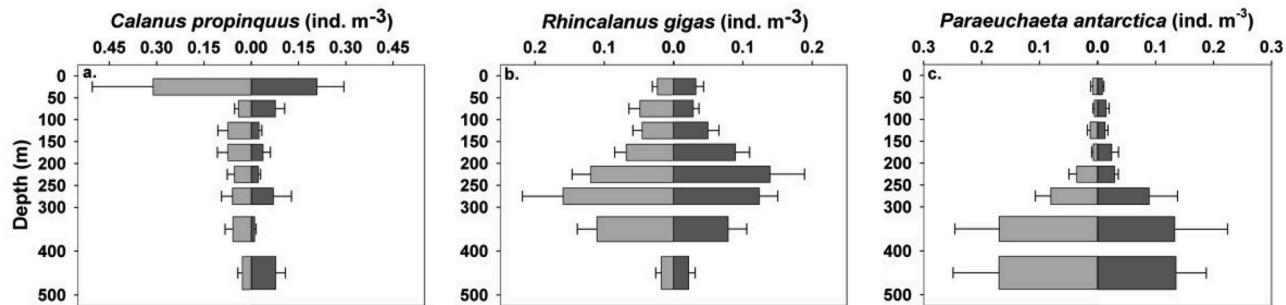


Fig. 4. Mean day (light gray, left) and night (dark gray, right) abundance of the calanoid copepods *Calanus propinquus* (a), *Rhincalanus gigas* (b), and *Paraeuchaeta antarctica* (c) sampled at discrete depth intervals from 0–500 m for the full shelf sampling region. Error bars indicate one standard error. Full $n = 12$ –17. Note different scaling on x-axes.

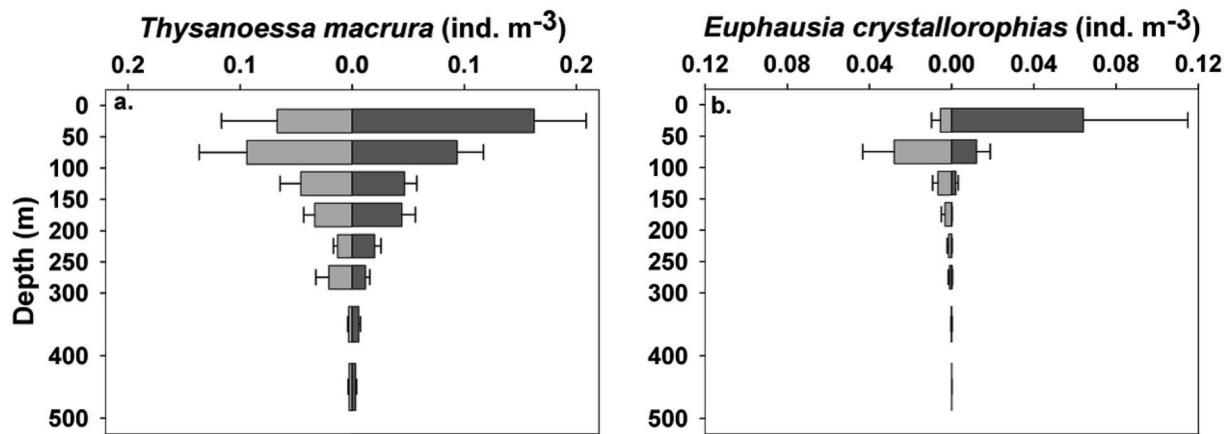


Fig. 5. Mean day (light gray, left) and night (dark gray, right) abundance of the euphausiids *Thysanoessa macrura* (a) and *Euphausia crystallorophias* (b) sampled at discrete depth intervals from 0–500 m for the full shelf sampling region. Error bars indicate one standard error. Full $n = 17$ –22. Note different scaling on x-axes.

and Hewitt, 1995; Ross et al., 1996). The daytime formation of larger schools and nighttime dispersal into smaller schools may be a more consistent predator avoidance behavior for *E. superba* in the summer (Everson, 1983; Zhou and Dorland, 2004; Tarling et al., 2018).

4.2.2. Mesopelagic carnivores and detritivores

The copepod *P. antarctica*, ostracods, chaetognaths, and *Tomopteris* spp. polychaetes were concentrated in the mesopelagic zone regardless of the diel cycle, and together compose a deep carnivorous and detritivorous assemblage. The carnivorous *P. antarctica* (synonym *Euchaeta antarctica*) preyed mainly on other copepods in the Weddell Sea

Table 3

Diel, depth-integrated zooplankton abundance (0–120 m) from grid-wide epipelagic tows across the PAL LTER sampling region from 1993–2017. n varies because not all taxa were identified consistently throughout the time series. p -values are for the Wilcoxon rank-sum test.

Taxon	Day (individuals m^{-2})			Night (individuals m^{-2})			Night:Day	p
	Mean	SE	n	Mean	SE	n		
Euphausiids								
<i>Thysanoessa macrura</i>	22.9	2.0	1063	36.9	6.8	197	1.6	0.0003
<i>Euphausia crystallorophias</i>	2.9	0.38	1067	6.2	2.3	196	2.1	0.77
<i>Euphausia superba</i>	13.7	2.0	1071	12.7	6.1	198	0.9	0.001
Other crustaceans								
Amphipoda	0.36	0.025	1026	0.75	0.13	185	2.1	2.6×10^{-12}
Gelatinous zooplankton								
<i>Salpa thompsoni</i>	4.9	1.4	1069	12.5	3.0	197	2.6	7.7×10^{-17}
<i>Limacina helicina antarctica</i>	7.3	0.62	1056	9.9	1.1	195	1.4	0.004
Gymnosomata	0.22	0.014	1050	0.31	0.039	193	1.4	0.0002
Chaetognatha	1.5	0.16	994	0.88	0.20	183	0.6	0.088
<i>Tomopteris</i> spp.	0.051	0.0049	966	0.095	0.014	181	1.9	0.0002

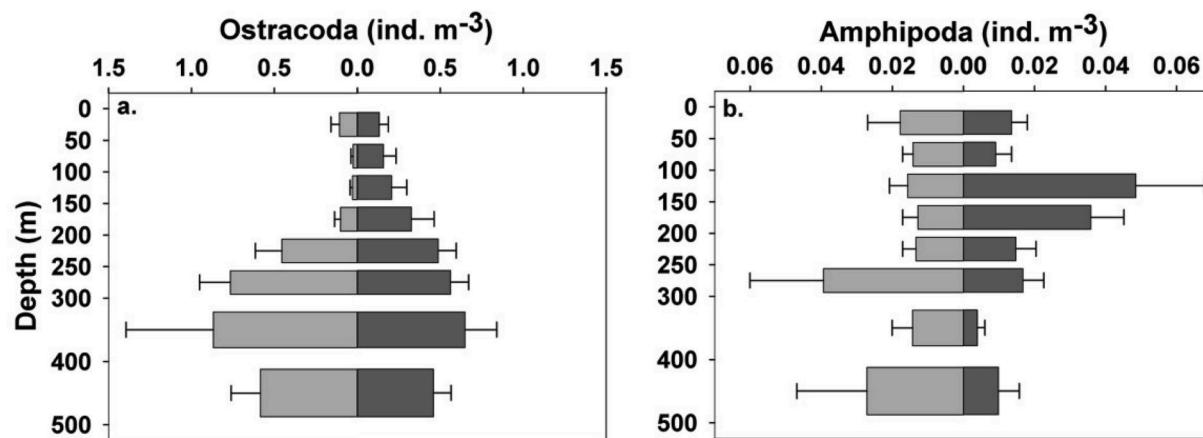


Fig. 6. Mean day (light gray, left) and night (dark gray, right) abundance of ostracods (a) and amphipods (b) sampled at discrete depth intervals from 0–500 m for the full shelf sampling region. Error bars indicate one standard error. Full n = 12–17. Note different scaling on x-axes.

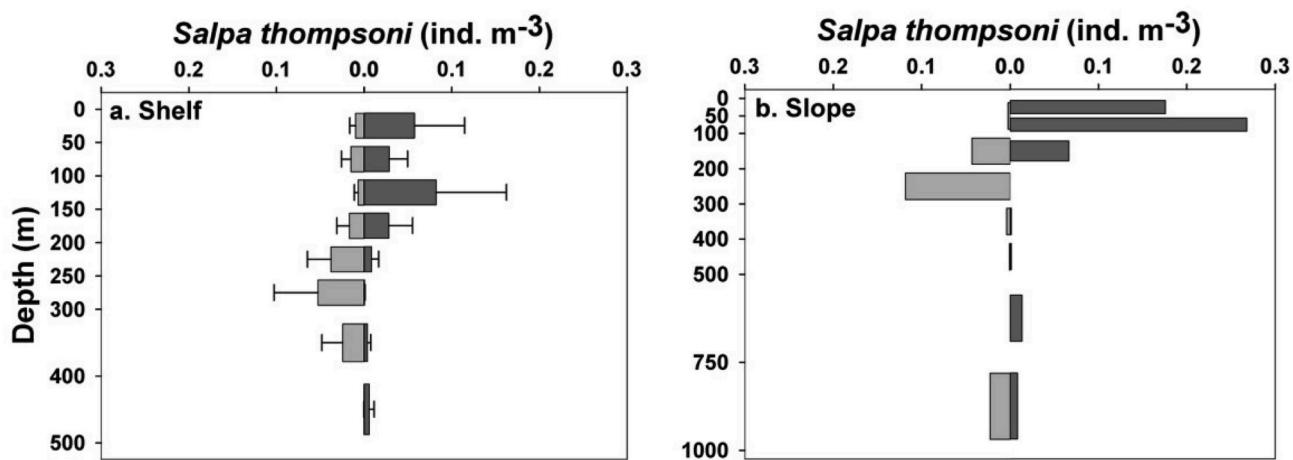


Fig. 7. Mean day (light gray, left) and night (dark gray, right) abundance of *Salpa thompsoni* from the full continental shelf sampling region (a) and a single pair of tows on the continental slope (b) sampled at discrete depth intervals from 0–500 m and 0–1000 m, respectively. Error bars indicate one standard error. Full Shelf n = 17–22; Slope n = 1. Note different scaling on y-axes.

(Hopkins and Torres, 1989) and near South Georgia where feeding continued through winter (Øresland and Ward, 1993). Thus, *P. antarctica* likely remains resident in the mesopelagic zone where metazoan prey is sufficiently abundant throughout the year. While vertically migrating ostracods feed in productive surface waters, the more numerous mesopelagic residents feed as carnivores or detritivores

(Angel, 1972; Lampitt et al., 1993; Vannier et al., 1998). Elevated ostracod abundance from 200–500 m was also observed in the northern WAP during summer (Blachowiak-Samolyk and Żmijewska, 1997) and in Marguerite Bay during autumn (Marrari et al., 2011). Chaetognaths along the WAP likely remained in deeper layers during summer to feed on abundant copepod prey as previously discussed. Chaetognaths were

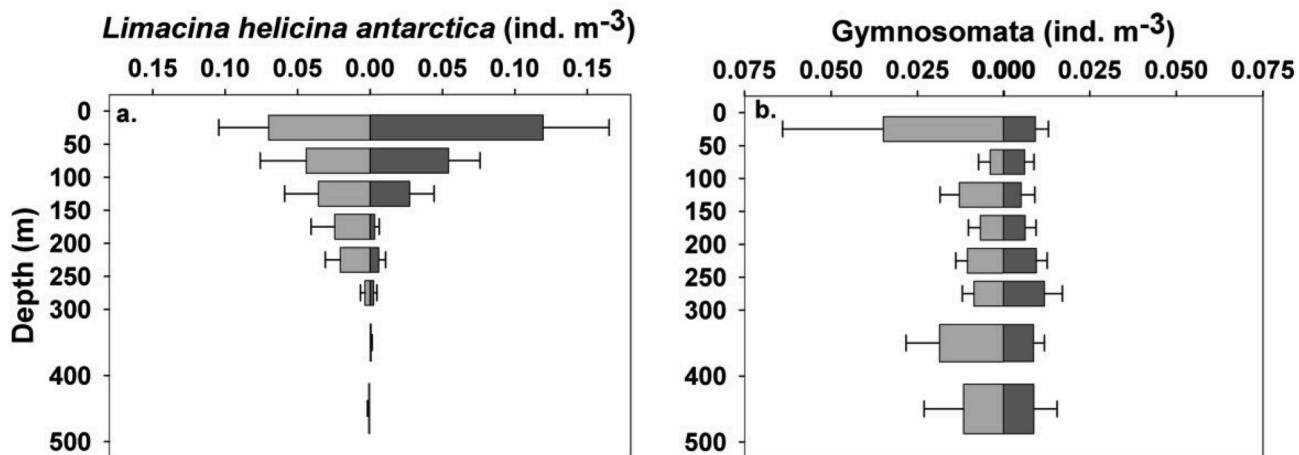


Fig. 8. Mean day (light gray, left) and night (dark gray, right) abundance of the pteropod *Limacina helicina antarctica* (a) and gymnosome pteropods (b) sampled at discrete depth intervals from 0–500 m for the full shelf sampling region. Error bars indicate one standard error. Full $n = 12\text{--}17$. Note different scaling on x-axes.

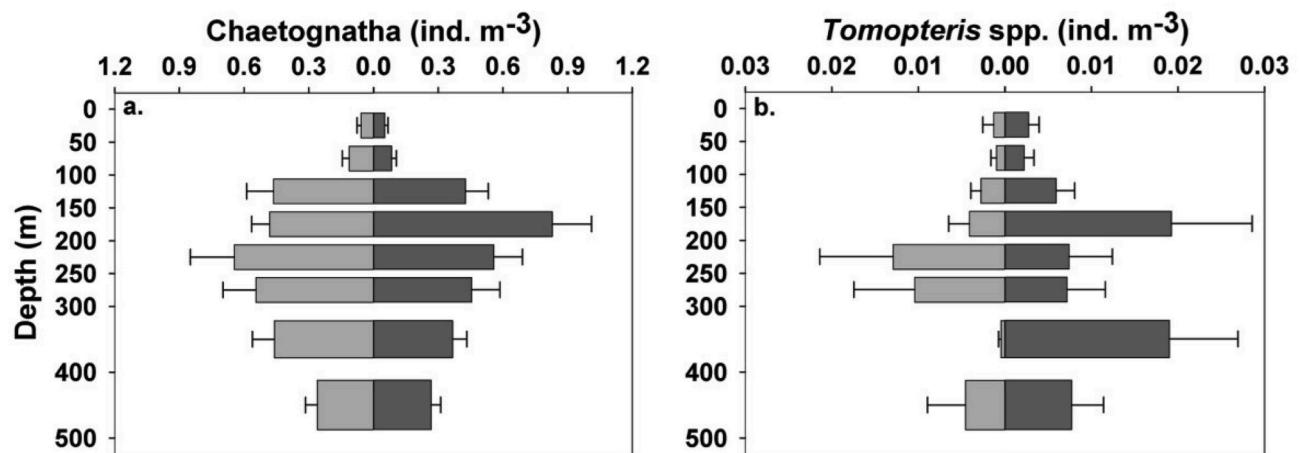


Fig. 9. Mean day (light gray, left) and night (dark gray, right) abundance of chaetognaths (a) and *Tomopteris* spp. polychaetes (b) sampled at discrete depth intervals from 0–500 m for the full shelf sampling region. Error bars indicate one standard error. Full $n = 12\text{--}17$. Note different scaling on x-axes.

Table 4

Statistics from multiple linear regression models assessing the impact of environmental variables on zooplankton ΔWMD from MOCNESS tows (0–500 m) along the WAP continental shelf from 2009–2015.

Variable	<i>n</i>	Coefficient	SE	<i>t</i>	<i>p</i>	Partial R ²
<i>Metridia gerlachei</i> ΔWMD (adjusted R ² = 0.76; <i>p</i> = 0.001)	11					
Photoperiod		-19.8	4.6	-4.3	0.003	0.70
Depth of chl- <i>a</i> maximum		2.27	0.47	4.8	0.001	0.75
Intercept		400.0	92.5	4.3	0.003	
<i>Ostracoda</i> ΔWMD (adjusted R ² = 0.79; <i>p</i> = 0.002)	10					
Photoperiod		-12.1	3.0	-4.1	0.005	0.70
Mixed layer depth		-0.666	0.23	-2.9	0.022	0.55
Intercept		283.8	59.7	4.8	0.002	

the numerically dominant macrozooplankton in the mesopelagic zone throughout the year in the Lazarev Sea (Flores et al., 2014) and are similarly important in the WAP mesopelagic zone. Small *Tomopteris* spp. polychaetes in the epipelagic zone fed on phytoplankton in the northern WAP during summer (Phleger et al., 1998) and during autumn in the Weddell Sea (Hopkins and Torres, 1989). However, *Tomopteris* spp. polychaetes in the mesopelagic zone are carnivores or detritivores

(Steinberg et al., 1994; Jumars et al., 2015). In particular, individuals larger than 20 mm are primarily carnivorous (Jumars et al., 2015), and large *Tomopteris* spp. individuals in our study exceeded 60 mm, further supporting their role as carnivores in the mesopelagic zone.

Amphipods and gymnosome pteropods were distributed throughout the water column, with a substantial portion of the population residing in the mesopelagic zone during day and night. A diet study in the Weddell Sea during March found multiple amphipod species were feeding mainly on copepods, larval euphausiids, and gelatinous zooplankton (Hopkins and Torres, 1989). Therefore, carnivory explains increased amphipod density below 100 m where metazoan prey is abundant. Although the highest gymnosome abundance was in surface waters, their consistent mesopelagic presence suggests gymnosomes consume other prey in addition to the shelled pteropod *L. h. antarctica*. A genetic diet analysis of the Arctic *Clione limacina* found this species fed on amphipods and calanoid copepods in addition to shelled pteropods (Kallevik, 2013). Similar to other mesopelagic zooplankton, amphipods and gymnosome pteropods likely have multiple feeding modes.

4.2.3. Seasonal vertical migrators

The copepods *M. gerlachei*, *R. gigas*, and *C. acutus* all make seasonal vertical migrations, although *M. gerlachei* and *R. gigas* feed through winter while *C. acutus* enters diapause at depth once it has acquired sufficient lipid reserves (Atkinson, 1998; Pasternak and Schnack-Schiel,

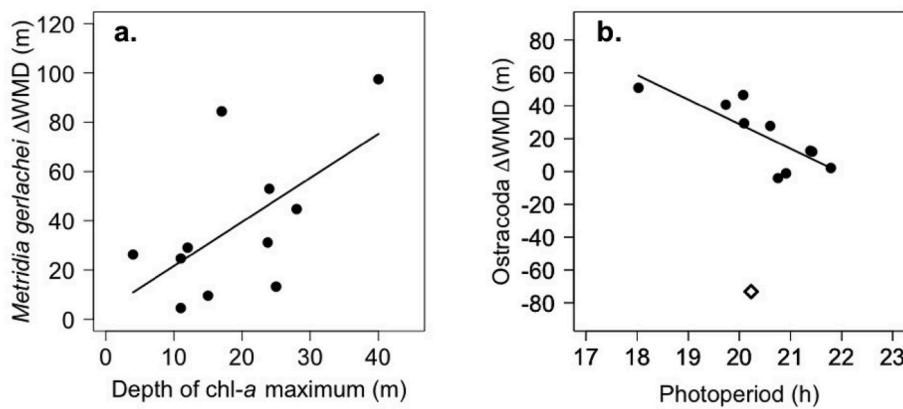


Fig. 10. Environmental controls on ΔWMD . (a) Depth of chlorophyll *a* maximum versus *Metridia gerlachei* ΔWMD . (b) Photoperiod versus Ostracod ΔWMD . Solid lines indicate the linear regression with all *M. gerlachei* data points and without the ostracod outlier value, indicated by an open diamond. *M. gerlachei*: $n = 11$, $p = 0.048$, $R^2 = 0.30$; Ostracod (without outlier): $n = 10$, $p = 0.005$, $R^2 = 0.60$.

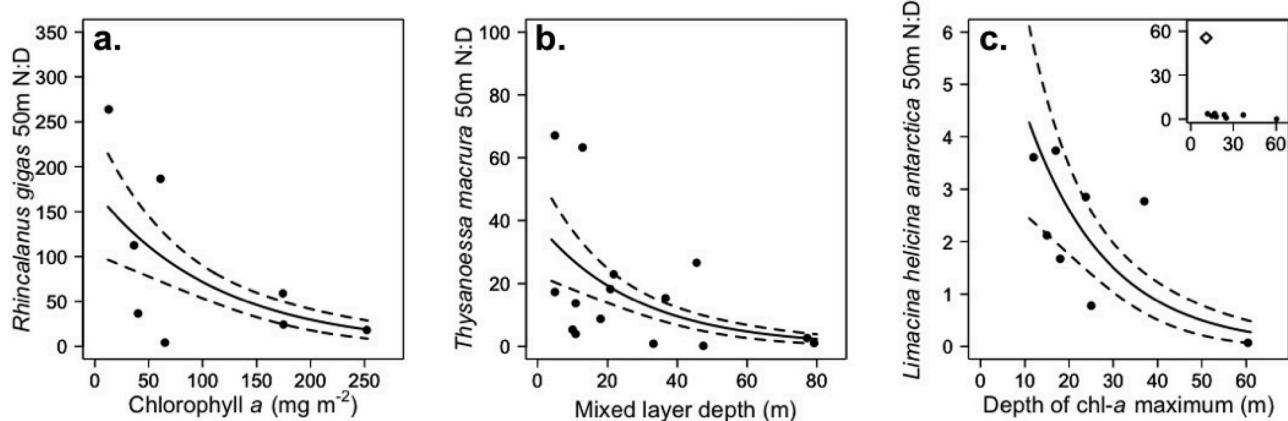


Fig. 11. Environmental controls on 50 m N:D. (a) Depth-integrated chlorophyll *a* concentration versus *Rhincalanus gigas* 50 m N:D. (b) Mixed layer depth versus *Thysanoessa macrura* 50 m N:D. (c) Depth of chlorophyll *a* maximum versus *Limacina helicina antarctica* 50 m N:D. Inset includes an outlier *L. h. antarctica* 50 m N:D value indicated by an open diamond. Solid line indicates generalized linear model fit. Dashed lines indicate one standard error. *R. gigas*: $n = 8$, $p = 0.031$, deviance explained = 35%; *T. macrura*: $n = 15$, $p = 0.008$, deviance explained = 27%; *L. h. antarctica* (without outlier): $n = 8$, $p = 0.035$, deviance explained = 40%.

2001; Schnack-Schiel, 2001). Elevated mesopelagic concentrations for these species in our study are more similar to autumn or winter depth distributions farther north in the Scotia Sea (Atkinson and Sinclair, 2000; Ward et al., 2012). High *M. gerlachei* and *R. gigas* concentrations from 200–500 m are likely indicative of carnivorous and detritivorous

feeding, which may be more important along the WAP where the productive season is shorter vs. lower latitudes. High mesopelagic abundance indicates *C. acutus* adults were likely in diapause and had not yet fully begun their ascent (Atkinson and Shreeve, 1995) in the Far South where we sampled pre-bloom conditions and mean depth-integrated chl-*a* was only 38 mg m^{-2} . Reduced seasonal sea ice coverage coincident with increasing phytoplankton biomass in the PAL LTER study area (Stammerjohn et al., 2008; Montes-Hugo et al., 2009) may result in earlier ascents for seasonally migrating copepods.

Indications that the pteropod *L. h. antarctica* conducts a seasonal vertical migration to feed in WAP surface waters during summer and overwinter at depth include that this species was concentrated from 0–100 m during our sampling but was most abundant from 100–200 m during autumn in Marguerite Bay (Marrari et al., 2011). Furthermore, *L. h. antarctica* is typically absent from a moored sediment trap sampling at 170 m on the WAP shelf during summer but commonly collected from June to October (Thibodeau et al., in review). A seasonal vertical migration for the closely related *Limacina helicina helicina* is also suggested in the Arctic Ocean (Kobayashi, 1974).

4.3. Environmental controls on DVM

4.3.1. DVM between epipelagic and mesopelagic zones

The copepod *M. gerlachei* and ostracods made shorter DVMs as photoperiod increased from 17 to 22 h, consistent with previous work demonstrating the sensitivity of polar zooplankton to seasonal

Table 5

Statistics from generalized linear models (with a gamma distribution and log link function) assessing the impact of environmental variables on zooplankton 50 m N:D from MOCNESS tows along the WAP continental shelf from 2009–2015 (for *Rhincalanus gigas* and *Limacina helicina antarctica*) and 2009–2017 (for *Thysanoessa macrura*).

Variable	<i>n</i>	Coefficient	SE	<i>t</i>	<i>p</i>
<i>Rhincalanus gigas</i> 50 m N:D (deviance explained = 35%)	8				
Chl- <i>a</i> concentration		-0.0088	0.0031	-2.8	0.031
Intercept		5.15	0.41	12.6	1.5×10^{-5}
<i>Thysanoessa macrura</i> 50 m N:D (deviance explained = 27%)	15				
Mixed layer depth		-0.035	0.011	-3.1	0.008
Intercept		3.67	0.42	8.7	8.8×10^{-7}
<i>Limacina helicina antarctica</i> 50 m N:D (deviance explained = 40%)	8				
Depth of chl- <i>a</i> maximum		-0.055	0.020	-2.7	0.035
Intercept		2.06	0.61	3.4	0.015

irradiance cycles. Acoustic studies throughout the Southern Ocean (64°S to 74°S) suggest that DVM ceases during summer (Cisewski et al., 2010; Cisewski and Strass, 2016; Picco et al., 2017). In these studies, sound scattering layers remained in surface waters around-the-clock rather than descending during the day. However, our results show some taxa continue to migrate between the epipelagic and mesopelagic zones, responding to relatively small changes in photoperiod during Antarctic summer from 64°S to 70°S .

Metridia gerlachei made DVMs over a greater depth range when Z_{SCM} was deeper, likely due to changing phytoplankton availability and predation risk. Deeper Z_{SCM} was associated with reduced light attenuation. With a deep Z_{SCM} , *M. gerlachei* encountered maximum phytoplankton concentrations farther from the surface at night, but DVM distance increased as *M. gerlachei* migrated deeper to avoid visual predators during day. This finding is consistent with previous work in the southern California Current, where reduced light attenuation was associated with longer DVM distances – particularly deeper daytime depths – for migrating copepods (Ohman and Romagnan, 2016). *Metridia gerlachei* (mean prosome length 3 mm; Gleiber, 2014) is within the size range of the strongest vertical migrators in that study, and therefore may be similarly susceptible to visual predators. Z_{SCM} thus influences *M. gerlachei* DVM distance directly by concentrating prey distribution and indirectly by modulating predation threat (i.e., via light attenuation).

Ostracods made shorter DVMs when MLD was deeper, likely because migrating ostracods ascended from the mesopelagic zone at night until reaching elevated phytoplankton concentrations in the mixed layer. A shallow mixed layer results in a longer nighttime feeding ascent and a longer return to mesopelagic daytime residence depth. In an acoustic study during Arctic autumn, migrating zooplankton sound scattering layers were coincident with, or just below, the MLD at midnight (Berge et al., 2014). Deeper MLD is associated with reduced sea ice coverage along the WAP (Schofield et al., 2018), which may result in shorter ostracod DVMs under future regional climate conditions.

4.3.2. DVM within epipelagic zone

Depth-integrated chlorophyll *a* concentration influenced DVM by the copepod *R. gigas* within the epipelagic zone. *Rhincalanus gigas* remained in the surface 0–50 m during day and night to feed on elevated phytoplankton biomass, as indicated by decreasing N:D with increasing chl-*a*. Gut content analysis showed *R. gigas* feeds primarily on phytoplankton during summer (Pasternak and Schnack-Schiel, 2001). Elevated chl-*a* was also associated with increased light attenuation, reducing the susceptibility of *R. gigas* to visual predators in surface waters and limiting any benefit gained by daytime migration out of the upper 50 m. Future long-term declines in regional sea ice coverage, upper ocean stability, and chl-*a* concentration in the PAL LTER study region (Montes-Hugo et al., 2009; Brown et al., 2019) may increase the amplitude of *R. gigas* DVM.

The 0–50 m N:D ratio for the euphausiid *T. macrura* and for the pteropod *L. h. antarctica* increased with shallower MLD and Z_{SCM} , respectively, as these taxa appeared to cue on vertical phytoplankton distribution. MLD and Z_{SCM} were positively correlated in our study. We suggest increased 0–50 m N:D indicates a larger portion of the population migrated into the upper 50 m at night when phytoplankton was concentrated near the surface. When MLD and Z_{SCM} were deeper, *T. macrura* and *L. h. antarctica* DVM into the upper 50 m likely decreased because phytoplankton availability and predator avoidance were both maximized below the 50 m threshold of our sampling resolution. During autumn, Arctic zooplankton sound scattering layers migrated to the MLD at midnight (Berge et al., 2014). Migrating pteropods and copepods ascended to the Z_{SCM} at night during Arctic summer (Daase et al., 2016). Deeper MLD under reduced sea ice conditions along the WAP (Schofield et al., 2018) may result in deeper nighttime distributions for *T. macrura* euphausiids and *L. h. antarctica* pteropods.

4.4. Zooplankton-mediated carbon export

Zooplankton vertical structure and behavior play key roles in mediating carbon export (Steinberg and Landry, 2017; Cavan et al., 2019), and regional, taxon-specific data are needed to accurately model the contribution of zooplankton DVM to the global biological carbon pump (e.g., Aumont et al., 2018; Archibald et al., 2019). DVM between the epipelagic and mesopelagic zones by WAP zooplankton through Antarctic summer likely results in substantial active carbon transport out of the euphotic zone, which may help resolve surprisingly low regional particle export to primary production ratios (Stukel et al., 2015; Ducklow et al., 2018). Zooplankton DVM is an important control on POC export in the Scotia Sea (Cavan et al., 2015; Liszka et al., 2019), and likely plays a similar role along the WAP where zooplankton fecal pellets constitute 67% of summer POC flux at 170 m (Gleiber et al., 2012). Future work also should estimate active transport of dissolved carbon (i.e., respiration of CO₂ and excretion of DOC at depth). Additionally, seasonal vertical migrators (e.g., *C. acutus* copepods and *L. h. antarctica* pteropods) transfer carbon to the ocean interior as they respire and die in the mesopelagic zone, but this “lipid pump” (for copepods) is yet to be quantified in the Southern Ocean (Kobari et al., 2008; Jónasdóttir et al., 2015). Abundant mesopelagic zooplankton also consume sinking detritus and produce fecal pellets, which regulates POC availability to mesopelagic and benthic food webs (Wilson et al., 2008; Belcher et al., 2017). Our findings show it will be essential to consider species feeding ecology and variable DVM amplitudes when assessing zooplankton roles in Southern Ocean carbon cycling. Finally, documented long-term changes in WAP zooplankton composition (e.g., Steinberg et al., 2015; Thibodeau et al., 2019) as well as phytoplankton biomass and vertical distribution (e.g., Brown et al., 2019) will alter zooplankton-mediated export pathways.

Author contributions

John A. Conroy: Conceptualization, Investigation, Formal analysis, Visualization, Writing – original draft. **Deborah K. Steinberg:** Conceptualization, Methodology, Investigation, Writing – review & editing, Funding acquisition. **Patricia S. Thibodeau:** Conceptualization, Investigation, Writing – review & editing. **Oscar Schofield:** Conceptualization, Investigation, Writing – review & editing.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

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References

- Aksnes, D.L., Giske, J., 1993. A theoretical model of aquatic visual feeding. *Ecol. Model.* 67, 233–250. [https://doi.org/10.1016/0304-3800\(93\)90007-F](https://doi.org/10.1016/0304-3800(93)90007-F).
- Andersen, V., Guanova, A., Nival, P., Ruellet, T., 2001. Zooplankton community during the transition from spring bloom to oligotrophy in the open NW mediterranean and effects of wind events. 2. Vertical distributions and migrations. *J. Plankton Res.* 23, 243–261. <https://doi.org/10.1093/plankt/23.3.243>.
- Angel, M.V., 1979. Studies on Atlantic halocyprid ostracods: their vertical distributions and community structure in the central gyre region along latitude 30°N from off Africa to Bermuda. *Prog. Oceanogr.* 8, 3–124. [https://doi.org/10.1016/0079-6611\(79\)90009-0](https://doi.org/10.1016/0079-6611(79)90009-0).
- Angel, M.V., 1972. Planktonic oceanic ostracods—historical, present and future. *Proc. R. Soc. Edinb. Sect. B Biol. Sci.* 73, 213–228. <https://doi.org/10.1017/S0080455X00002277>.
- Archibald, K.M., Siegel, D.A., Doney, S.C., 2019. Modeling the impact of zooplankton diel vertical migration on the carbon export flux of the biological pump. *Global Biogeochem. Cycles* 33, 181–199. <https://doi.org/10.1029/2018GB005983>.
- Atkinson, A., 1998. Life cycle strategies of epipelagic copepods in the Southern Ocean. *J. Mar. Syst.* 15, 289–311. [https://doi.org/10.1016/S0924-7963\(97\)00081-X](https://doi.org/10.1016/S0924-7963(97)00081-X).
- Atkinson, A., Shreeve, R.S., 1995. Response of the copepod community to a spring bloom in the Bellingshausen Sea. *Deep Sea Res. Part II Top. Stud. Oceanogr.* 42, 1291–1311. [https://doi.org/10.1016/0967-0645\(95\)00057-W](https://doi.org/10.1016/0967-0645(95)00057-W).
- Atkinson, A., Siegel, V., Pakhomov, E.A., Rothery, P., Loeb, V., Ross, R.M., Quetin, L.B., Schmidt, K., Fretwell, P., Murphy, E.J., Tarling, G.A., Fleming, A.H., 2008. Oceanic circumpolar habitats of Antarctic krill. *Mar. Ecol. Prog. Ser.* 362, 1–23. <https://doi.org/10.3354/meps07498>.
- Atkinson, A., Sinclair, J.D., 2000. Zonal distribution and seasonal vertical migration of copepod assemblages in the Scotia Sea. *Polar Biol.* 23, 46–58. <https://doi.org/10.1007/s003000050007>.
- Atkinson, A., Ward, P., Williams, R., Poulet, S.A., 1992a. Diel vertical migration and feeding of copepods at an oceanic site near South Georgia. *Mar. Biol.* 113, 583–593. <https://doi.org/10.1007/BF00349702>.
- Atkinson, A., Ward, P., Williams, R., Poulet, S.A., 1992b. Feeding rates and diel vertical migration of copepods near South Georgia: comparison of shelf and oceanic sites. *Mar. Biol.* 114, 49–56. <https://doi.org/10.1007/BF00350855>.
- Aumont, O., Maury, O., Lefort, S., Bopp, L., 2018. Evaluating the potential impacts of the diurnal vertical migration by marine organisms on marine biogeochemistry. *Global Biogeochem. Cycles* 32, 1622–1643. <https://doi.org/10.1029/2018GB005886>.
- Bartoň, K., 2016. MuMIn: multi-model inference. R package version 1.15.6. <https://CRAN.R-project.org/package=MuMIn>.
- Belcher, A., Manno, C., Ward, P., Henson, S.A., Sanders, R., Tarling, G.A., 2017. Copepod faecal pellet transfer through the meso- and bathypelagic layers in the Southern Ocean in spring. *Biogeosciences* 14 (6), 1511–1525. <https://doi.org/10.5194/bg-14-1511-2017>.
- Berge, J., Cottier, F., Varpe, Ø., Renaud, P.E., Falk-Petersen, S., Kwasniewski, S., Griffiths, C., Sørøide, J.E., Johnsen, G., Aubert, A., Bjørke, O., Hovinen, J., Jung-Madsen, S., Tveit, M., Majaneva, S., 2014. Arctic complexity: a case study on diel vertical migration of zooplankton. *J. Plankton Res.* 36, 1279–1297. <https://doi.org/10.1093/plankt/fbu059>.
- Blachowiak-Samolyk, K., Kwasniewski, S., Richardson, K., Dmoch, K., Hansen, E., Hop, H., Falk-Petersen, S., Mouritsen, L.T., 2006. Arctic zooplankton do not perform diel vertical migration (DVM) during periods of midnight sun. *Mar. Ecol. Prog. Ser.* 308, 101–116. <https://doi.org/10.3354/meps308101>.
- Blachowiak-Samolyk, K., Źmijewska, M.I., 1997. Planktonic ostracoda in Croker Passage (Antarctic Peninsula) during two austral seasons: summer 1985/1986 and winter 1989. *Pol. Polar Res.* 18, 79–87.
- Brown, M.S., Munro, D.R., Feehan, C.J., Sweeney, C., Ducklow, H.W., Schofield, O.M., 2019. Enhanced oceanic CO₂ uptake along the rapidly changing West Antarctic Peninsula. *Nat. Clim. Change* 9, 678–683. <https://doi.org/10.1038/s41558-019-0552-3>.
- Carvalho, F., Kohut, J., Oliver, M.J., Schofield, O., 2017. Defining the ecologically relevant mixed-layer depth for Antarctica's coastal seas. *Geophys. Res. Lett.* 44, 338–345. <https://doi.org/10.1002/2016GL071205>.
- Casareto, B.E., Nemoto, T., 1986. Salps of the Southern Ocean (Australian sector) during the 1983–84 summer, with special reference to the species *Salpa thompsoni*, Foxtoun 1961. *Mem. Natl. Inst. Polar Res.* 40, 221–239.
- Cavan, E.L., Laurenceau-Cornec, E.C., Bressac, M., Boyd, P.W., 2019. Exploring the ecology of the mesopelagic biological pump. *Prog. Oceanogr.* 176, 102125. <https://doi.org/10.1016/j.pocean.2019.102125>.
- Cavan, E.L., Le Moigne, F.A.C., Poulton, A.J., Tarling, G.A., Ward, P., Daniels, C.J., Fragoso, G.M., Sanders, R.J., 2015. Attenuation of particulate organic carbon flux in the Scotia Sea, Southern Ocean, is controlled by zooplankton fecal pellets. *Geophys. Res. Lett.* 42, 821–830. <https://doi.org/10.1002/2014GL062744>.
- Cisewski, B., Strass, V.H., 2016. Acoustic insights into the zooplankton dynamics of the eastern Weddell Sea. *Prog. Oceanogr.* 144, 62–92. <https://doi.org/10.1016/j.pocean.2016.03.005>.
- Cisewski, B., Strass, V.H., Rhein, M., Krägesky, S., 2010. Seasonal variation of diel vertical migration of zooplankton from ADCP backscatter time series data in the Lazarev Sea, Antarctica. *Deep-Sea Res. Part I Oceanogr. Res. Pap.* 57, 78–94. <https://doi.org/10.1016/j.dsr.2009.10.005>.
- Cohen, J.H., Forward Jr., R.B., 2009. Zooplankton diel vertical migration – a review of proximate control. *Oceanogr. Mar. Biol. Annu. Rev.* 47, 77–109. <https://doi.org/10.1201/9781420094220.ch2>.
- Cottier, F.R., Tarling, G.A., Wold, A., Falk-Petersen, S., 2006. Unsynchronised and synchronised vertical migration of zooplankton in a high Arctic fjord. *Limnol. Oceanogr.* 51, 2586–2599. <https://doi.org/10.4319/lo.2006.51.6.2586>.
- Daase, M., Hop, H., Falk-Petersen, S., 2016. Small-scale diel vertical migration of zooplankton in the High Arctic. *Polar Biol.* 39, 1213–1223. <https://doi.org/10.1007/s00300-015-1840-7>.
- Demer, D.A., Hewitt, R.P., 1995. Bias in acoustic biomass estimates of *Euphausia superba* due to diel vertical migration. *Deep-Sea Res. Part I Oceanogr. Res. Pap.* 42, 455–475. [https://doi.org/10.1016/0967-0637\(94\)E0005-C](https://doi.org/10.1016/0967-0637(94)E0005-C).
- Ducklow, H., Clarke, A., Dickhut, R., Doney, S.C., Geisz, H., Huang, K., Martinson, D.G., Meredith, M.P., Moeller, H.V., Montes-Hugo, M., Schofield, O., Stammerjohn, S.E., Steinberg, D., Fraser, W., 2012. The Marine System of the Western Antarctic Peninsula. In: Rogers, A.D., Johnston, N.M., Murphy, E.J., Clarke, A. (Eds.), *Antarctic Ecosystems*. John Wiley & Sons, Ltd, pp. 121–159.
- Ducklow, H.W., Stukel, M.R., Eveleth, R., Doney, S.C., Jickells, T., Schofield, O., Baker, A.R., Brindle, J., Chance, R., Cassar, N., 2018. Spring–summer net community production, new production, particle export and related water column biogeochemical processes in the marginal sea ice zone of the Western Antarctic Peninsula 2012–2014. *Philos. Trans. R. Soc. A* 376, 20170177. <https://doi.org/10.1098/rsta.2017.0177>.
- Everson, I., 1987. Some aspects of the small scale distribution of *Euphausia crystallorophias*. *Polar Biol.* 8, 9–15. <https://doi.org/10.1007/BF00297158>.
- Everson, I., 1983. Variations in vertical distribution and density of krill swarms in the vicinity of South Georgia. *Mem. Natl. Inst. Polar Res.* 27, 84–92.
- Falk-Petersen, S., Leu, E., Berge, J., Kwasniewski, S., Nygård, H., Røstad, A., Keskinen, E., Thormar, J., von Quillfeldt, C., Wold, A., Gulliksen, B., 2008. Vertical migration in high Arctic waters during autumn 2004. *Deep Sea Res. Part II Top. Stud. Oceanogr.* 55, 2275–2284. <https://doi.org/10.1016/j.dsr2.2008.05.010>.
- Flores, H., Hunt, B.P.V., Kruse, S., Pakhomov, E.A., Siegel, V., van Franeker, J.A., Strass, V., Van de Putte, A.P., Meesters, E.H.W.G., Bathmann, U., 2014. Seasonal changes in the vertical distribution and community structure of Antarctic macrozooplankton and microneuston. *Deep Sea Res. Part I Oceanogr. Res. Pap.* 84, 127–141. <https://doi.org/10.1016/j.dsr.2013.11.001>.
- Fortier, M., Fortier, L., Hattori, H., Saito, H., Legendre, L., 2001. Visual predators and the diel vertical migration of copepods under Arctic sea ice during the midnight sun. *J. Plankton Res.* 23, 1263–1278. <https://doi.org/10.1093/plankt/23.11.1263>.
- Gleiber, M.R., 2014. Long-term Change in Copepod Community Structure in the Western Antarctic Peninsula: Linkage to Climate and Implications for Carbon Cycling. Master's Thesis The College of William & Mary.
- Gleiber, M.R., Steinberg, D.K., Ducklow, H.W., 2012. Time series of vertical flux of zooplankton fecal pellets on the continental shelf of the western Antarctic Peninsula. *Mar. Ecol. Prog. Ser.* 471, 23–36. <https://doi.org/10.3354/meps10021>.
- Gliwicz, M.Z., 1986. Predation and the evolution of vertical migration in zooplankton. *Nature* 320, 746–748. <https://doi.org/10.1038/320746a0>.
- Godelevska, M., Klusek, Z., 1987. Vertical distribution and diurnal migrations of krill — *Euphausia superba* Dana — from hydroacoustical observations, SIBEX. *Polar Biol.* 8, 17–22. <https://doi.org/10.1007/BF00297159>.
- Hays, G.C., 2003. A review of the adaptive significance and ecosystem consequences of zooplankton diel vertical migrations. In: *Migrations and Dispersal of Marine Organisms*. Springer, pp. 163–170. https://doi.org/10.1007/978-94-017-2276-6_18.
- Hobbs, L., Cottier, F.R., Last, K.S., Berge, J., 2018. Pan-Arctic diel vertical migration during the polar night. *Mar. Ecol. Prog. Ser.* 605, 61–72. <https://doi.org/10.3354/meps12753>.
- Hopkins, T.L., 1985. The zooplankton community of Croker Passage, Antarctic Peninsula. *Polar Biol.* 4, 161–170. <https://doi.org/10.1007/BF00263879>.
- Hopkins, T.L., Torres, J.J., 1989. Midwater food web in the vicinity of a marginal ice zone in the western Weddell Sea. *Deep-Sea Res. Part I Oceanogr. Res. Pap.* 36, 543–560. [https://doi.org/10.1016/0198-0149\(89\)90005-8](https://doi.org/10.1016/0198-0149(89)90005-8).
- Hunt, B.P.V., Pakhomov, E.A., Hosie, G.W., Siegel, V., Ward, P., Bernard, K., 2008. Pteropods in Southern Ocean ecosystems. *Prog. Oceanogr.* 78, 193–221. <https://doi.org/10.1016/j.pocean.2008.06.001>.
- Huntley, M., Escritor, F., 1991. Dynamics of *Calanoides acutus* (Copepoda: Calanoida) in Antarctic coastal waters. *Deep-Sea Res. Part I Oceanogr. Res. Pap.* 38, 1145–1167. [https://doi.org/10.1016/0198-0149\(91\)90100-T](https://doi.org/10.1016/0198-0149(91)90100-T).
- Hurvich, C.M., Tsai, C.-L., 1989. Regression and time series model selection in small samples. *Biometrika* 76, 297–307. <https://doi.org/10.1093/biomet/76.2.297>.
- Jónasdóttir, S.H., Visser, A.W., Richardson, K., Heath, M.R., 2015. Seasonal copepod lipid pump promotes carbon sequestration in the deep North Atlantic. *Proc. Natl. Acad. Sci. Unit. States Am.* 112, 12122–12126. <https://doi.org/10.1073/pnas.1512110112>.

- Jumars, P.A., Dorgan, K.M., Lindsay, S.M., 2015. Diet of worms emended: an update of polychaete feeding guilds. Annu. Rev. Mar. Sci. 7, 497–520. <https://doi.org/10.1146/annurev-marine-010814-020007>.
- Kallevik, L.H.F., 2013. Alternative prey choice in the pteropod *Clione limacina* (Gastropoda) studied by DNA-based methods.. Master's Thesis UIT Norges arktiske universitet.
- King, A., LaCasella, E.L., 2003. Seasonal variations in abundance, diel vertical migration, and population structure of *Metridia gerlachei* at Port Foster, Deception Island, Antarctica. Deep Sea Res. Part II Top. Stud. Oceanogr. 50, 1753–1763. [https://doi.org/10.1016/S0967-0645\(03\)00091-2](https://doi.org/10.1016/S0967-0645(03)00091-2).
- Kirk, J.T.O., 2011. Light and Photosynthesis in Aquatic Ecosystems, third ed. Cambridge University Press, Cambridge.
- Klevjer, T.A., Irigoien, X., Røstad, A., Fraile-Nuez, E., Benítez-Barrios, V.M., Kaartvedt, S., 2016. Large scale patterns in vertical distribution and behaviour of mesopelagic scattering layers. Sci. Rep. 6, srep19873 <https://doi.org/10.1038/srep19873>.
- Kobari, T., Steinberg, D.K., Ueda, A., Tsuda, A., Silver, M.W., Kitamura, M., 2008. Impacts of ontogenetically migrating copepods on downward carbon flux in the western subarctic Pacific Ocean. Deep Sea Res. Part II Top. Stud. Oceanogr. 55, 1648–1660. <https://doi.org/10.1016/j.dsrt.2008.04.016>.
- Kobayashi, H.A., 1974. Growth cycle and related vertical distribution of the thecosomatous pteropod *Spiratella ("Limicina") helicina* in the central Arctic Ocean. Mar. Biol. 26, 295–301. <https://doi.org/10.1007/BF00391513>.
- La, H.S., Kang, M., Dahms, H.-U., Ha, H.K., Yang, E.J., Lee, H., Kim, Y.N., Chung, K.H., Kang, S.-H., 2015a. Characteristics of mesozooplankton sound-scattering layer in the Pacific Summer Water, Arctic Ocean. Deep Sea Res. Part II Top. Stud. Oceanogr. 120, 114–123. <https://doi.org/10.1016/j.dsrt.2015.01.005>.
- La, H.S., Lee, H., Fielding, S., Kang, D., Ha, H.K., Atkinson, A., Park, J., Siegel, V., Lee, S., Shin, H.C., 2015b. High density of ice krill (*Euphausia crystallorophias*) in the Amundsen Sea coastal polynya, Antarctica. Deep-Sea Res. Part I Oceanogr. Res. Pap. 95, 75–84. <https://doi.org/10.1016/j.dsr.2014.09.002>.
- Lalli, C.M., Gilmer, R.W., 1989. Pelagic Snails: the Biology of Holoplanktonic Gastropod Mollusks. Stanford University Press.
- Lampitt, R.S., Wishner, K.F., Turley, C.M., Angel, M.V., 1993. Marine snow studies in the Northeast Atlantic Ocean: distribution, composition and role as a food source for migrating plankton. Mar. Biol. 116, 689–702. <https://doi.org/10.1007/BF00355486>.
- Liszka, C.M., Manno, C., Stowasser, G., Robinson, C., Tarling, G.A., 2019. Mesozooplankton community composition controls fecal pellet flux and remineralization depth in the Southern Ocean. Front. Mar. Sci. 6 <https://doi.org/10.3389/fmars.2019.00230>.
- Loeb, V.J., Shulенberger, E., 1987. Vertical distributions and relations of euphausiid populations off Elephant Island, March 1984. Polar Biol. 7, 363–373. <https://doi.org/10.1007/BF00293226>.
- Lopez, M.D.G., Huntley, M.E., 1995. Feeding and diel vertical migration cycles of *Metridia gerlachei* (Giesbrecht) in coastal waters of the Antarctic Peninsula. Polar Biol. 15, 21–30.
- Ludvigsen, M., Berge, J., Geoffroy, M., Cohen, J.H., De La Torre, P.R., Nornes, S.M., Singh, H., Sørensen, A.J., Daase, M., Johnsen, G., 2018. Use of an Autonomous Surface Vehicle reveals small-scale diel vertical migrations of zooplankton and susceptibility to light pollution under low solar irradiance. Sci. Adv. 4, eaap9887 <https://doi.org/10.1126/sciadv.aap9887>.
- Marrari, M., Daly, K.L., Timonin, A., Semenova, T., 2011. The zooplankton of Marguerite Bay, western Antarctic Peninsula—Part II: vertical distributions and habitat partitioning. Deep Sea Res. Part II Top. Stud. Oceanogr. 58, 1614–1629. <https://doi.org/10.1016/j.dsrt.2010.12.006>.
- Martinson, D.G., Stammerjohn, S.E., Iannuzzi, R.A., Smith, R.C., Vernet, M., 2008. Western Antarctic Peninsula physical oceanography and spatio-temporal variability. Deep Sea Res. Part II Top. Stud. Oceanogr. 55, 1964–1987. <https://doi.org/10.1016/j.dsrt.2008.04.038>.
- Meeus, J.H., 1998. Astronomical Algorithms, second ed. Willmann-Bell, Incorporated, Richmond, VA.
- Montes-Hugo, M., Doney, S.C., Ducklow, H.W., Fraser, W., Martinson, D., Stammerjohn, S.E., Schofield, O., 2009. Recent changes in phytoplankton communities associated with rapid regional climate change along the western Antarctic Peninsula. Science 323, 1470–1473. <https://doi.org/10.1126/science.1164533>.
- Nordhausen, W., 1994b. Winter abundance and distribution of *Euphausia superba*, *E. crystallorophias*, and *Thysanoessa macrura* in Gerlache Strait and Crystal Sound, Antarctica. Mar. Ecol. Prog. Ser. 109, 131–142.
- Nordhausen, W., 1994a. Distribution and diel vertical migration of the euphausiid *Thysanoessa macrura* in Gerlache Strait, Antarctica. Polar Biol. 14, 219–229. <https://doi.org/10.1007/BF00239170>.
- Nordhausen, W., 1992. Distribution and growth of larval and adult *Thysanoessa macrura* (Euphausiaceae) in the Bransfield Strait Region, Antarctica. Mar. Ecol. Prog. Ser. 83, 185–196.
- Ohman, M.D., Frost, B.W., Cohen, E.B., 1983. Reverse diel vertical migration: an escape from invertebrate predators. Science 220, 1404–1407. <https://doi.org/10.1126/science.220.4604.1404>.
- Ohman, M.D., Romagnan, J.-B., 2016. Nonlinear effects of body size and optical attenuation on Diel Vertical Migration by zooplankton. Limnol. Oceanogr. 61, 765–770. <https://doi.org/10.1002/lno.10251>.
- Oresland, V., Ward, P., 1993. Summer and winter diet of four carnivorous copepod species around South Georgia. Mar. Ecol. Prog. Ser. 98, 73–78.
- Pakhomov, E.A., Hall, J., Williams, M.J.M., Hunt, B.P.V., Stevens, C.J., 2011. Biology of *Salpa thompsoni* in waters adjacent to the Ross Sea, Southern Ocean, during austral summer 2008. Polar Biol. 34, 257–271. <https://doi.org/10.1007/s00300-010-0878-9>.
- Parsons, R.T., Maita, Y., Lalli, C.M., 1984. A Manual of Chemical and Biological Methods for Seawater Analysis. Pergamon.
- Pasternak, A.F., Schnack-Schiel, S.B., 2001. Feeding patterns of dominant Antarctic copepods: an interplay of diapause, selectivity, and availability of food. Hydrobiologia 453, 25–36. <https://doi.org/10.1023/A:1013147413136>.
- Perissinotto, R., Pakhomov, E.A., 1998. The trophic role of the tunicate *Salpa thompsoni* in the Antarctic marine ecosystem. J. Mar. Syst. 17, 361–374. [https://doi.org/10.1016/S0924-7963\(98\)00049-9](https://doi.org/10.1016/S0924-7963(98)00049-9).
- Phleger, C.F., Nichols, P.D., Virtue, P., 1998. Lipids and trophodynamics of Antarctic zooplankton. Comp. Biochem. Physiol. B Biochem. Mol. Biol. 120, 311–323. [https://doi.org/10.1016/S0305-0491\(98\)10020-2](https://doi.org/10.1016/S0305-0491(98)10020-2).
- Piatkowski, U., 1985. Distribution, abundance and diurnal migration of macrozooplankton in Antarctic surface waters. Meeresforsch.-Rep. Mar. Res. 30, 264–279.
- Picco, P., Schiano, M.E., Pensieri, S., Bozzano, R., 2017. Time-frequency analysis of migrating zooplankton in the Terra Nova Bay polynya (Ross Sea, Antarctica). J. Mar. Syst. 166, 172–183. <https://doi.org/10.1016/j.jmarsys.2016.07.010>.
- R Core Team, 2016. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. <https://www.R-project.org>.
- Ringelberg, J., Van Gool, E., 2003. On the combined analysis of proximate and ultimate aspects in diel vertical migration (DVM) research. Hydrobiologia 491, 85–90. <https://doi.org/10.1023/A:1024407021957>.
- Ross, R.M., Quetin, L.B., Lascara, C.M., 1996. Distribution of Antarctic krill and dominant zooplankton west of the Antarctic Peninsula. Antarct. Res. 70, 199–217.
- Ross, R.M., Quetin, L.B., Martinson, D.G., Iannuzzi, R.A., Stammerjohn, S.E., Smith, R.C., 2008. Palmer LTER: patterns of distribution of five dominant zooplankton species in the epipelagic zone west of the Antarctic Peninsula, 1993–2004. Deep Sea Res. Part II Top. Stud. Oceanogr. 55, 2086–2105. <https://doi.org/10.1016/j.dsrt.2008.04.037>.
- Schnack-Schiel, S.B., 2001. Aspects of the study of the life cycles of Antarctic copepods. In: Copepods: Developments in Ecology, Biology and Systematics. Springer, pp. 9–24. https://doi.org/10.1007/0-306-47537-5_2.
- Schofield, O., Brown, M., Kohut, J., Nardelli, S., Saba, G., Waite, N., Ducklow, H., 2018. Changes in the upper ocean mixed layer and phytoplankton productivity along the West Antarctic Peninsula. Philos. Trans. R. Soc. A 376, 20170173. <https://doi.org/10.1098/rsta.2017.0173>.
- Stammerjohn, S.E., Martinson, D.G., Smith, R.C., Iannuzzi, R.A., 2008. Sea ice in the western Antarctic Peninsula region: spatio-temporal variability from ecological and climate change perspectives. Deep Sea Res. Part II Top. Stud. Oceanogr. 55, 2041–2058. <https://doi.org/10.1016/j.dsrt.2008.04.026>.
- Steemann Nielsen, E., 1952. The use of radio-active carbon (C^{14}) for measuring organic production in the sea. J. Conseil 18, 117–140.
- Steinberg, D.K., Cope, J.S., Wilson, S.E., Kobari, T., 2008. A comparison of mesopelagic mesozooplankton community structure in the subtropical and subarctic North Pacific Ocean. Deep Sea Res. Part II Top. Stud. Oceanogr. 55, 1615–1635. <https://doi.org/10.1016/j.dsrt.2008.04.025>.
- Steinberg, D.K., Landry, M.R., 2017. Zooplankton and the ocean carbon cycle. Annu. Rev. Mar. Sci. 9, 413–444. <https://doi.org/10.1146/annurev-marine-010814-015924>.
- Steinberg, D.K., Ruck, K.E., Gleiber, M.R., Garzio, L.M., Cope, J.S., Bernard, K.S., Stammerjohn, S.E., Martinson, D.G., Smith, R.C., Iannuzzi, R.A., 2008. Sea ice in the western Antarctic Peninsula region: spatio-temporal variability from ecological and climate change perspectives. Deep Sea Res. Part II Top. Stud. Oceanogr. 55, 2041–2058. <https://doi.org/10.1016/j.dsrt.2008.04.026>.
- Steinberg, D.K., Cope, J.S., Wilson, S.E., Kobari, T., 2008. A comparison of mesopelagic mesozooplankton community structure in the subtropical and subarctic North Pacific Ocean. Deep Sea Res. Part II Top. Stud. Oceanogr. 55, 1615–1635. <https://doi.org/10.1016/j.dsrt.2008.04.025>.
- Steinberg, D.K., Landry, M.R., 2017. Zooplankton and the ocean carbon cycle. Annu. Rev. Mar. Sci. 9, 413–444. <https://doi.org/10.1146/annurev-marine-010814-015924>.
- Steinberg, D.K., Ruck, K.E., Gleiber, M.R., Garzio, L.M., Cope, J.S., Bernard, K.S., Stammerjohn, S.E., Schofield, O.M.E., Quetin, L.B., Ross, R.M., 2015. Long-term (1993–2013) changes in macrozooplankton off the Western Antarctic Peninsula. Deep Sea Res. Part I Oceanogr. Res. Pap. 101, 54–70. <https://doi.org/10.1016/j.dsr.2015.02.009>.
- Steinberg, D.K., Silver, M.W., Pilskalm, C.H., Coale, S.L., Paduan, J.B., 1994. Midwater zooplankton communities on pelagic detritus (giant larvacean houses) in Monterey Bay, California. Limnol. Oceanogr. 39, 1606–1620. <https://doi.org/10.4319/lo.1994.39.7.1606>.
- Stich, H.-B., Lampert, W., 1981. Predator evasion as an explanation of diurnal vertical migration by zooplankton. Nature 293, 396–398. <https://doi.org/10.1038/293396a0>.
- Stukel, M.R., Asher, E., Couto, N., Schofield, O., Strebel, S., Tortell, P., Ducklow, H.W., 2015. The imbalance of new and export production in the Western Antarctic Peninsula, a potentially "leaky" ecosystem. Global Biogeochem. Cycles 29, 1400–1420. <https://doi.org/10.1002/2015GB005211>.
- Tarling, G.A., Thorpe, S.E., Fielding, S., Klevjer, T., Ryabov, A., Somerfield, P.J., 2018. Varying depth and swarm dimensions of open-ocean Antarctic krill *Euphausia superba* Dana, 1850 (Euphausiacea) over diel cycles. J. Crustac Biol. 38, 716–727. <https://doi.org/10.1093/jcbiol/ruy040>.
- Thibodeau, P.S., Steinberg, D.K., Stammerjohn, S.E., Hauri, C., 2019. Environmental controls on pteropod biogeography along the Western Antarctic Peninsula. Limnol. Oceanogr. 64, S240–S256. <https://doi.org/10.1002/lno.11041>.
- Thibodeau, P.S., Steinberg, D.K., McBride, C.E., Conroy, J.A., Keul, N., Ducklow, H.W., submitted. Long-term observations of pteropod phenology along the Western Antarctic Peninsula. Deep-Sea Res. Part I Oceanogr. Res. Pap.
- Van der Spoel, S., Dadon, J.R., 1999. Pteropoda. In: South Atlantic Zooplankton. Backhuys, pp. 868–1706.
- Vannier, J., Abe, K., Ikuta, K., 1998. Feeding in myodocapid ostracods: functional morphology and laboratory observations from videos. Mar. Biol. 132, 391–408. <https://doi.org/10.1007/s002270050406>.
- Ward, P., Atkinson, A., Tarling, G., 2012. Mesozooplankton community structure and variability in the Scotia Sea: a seasonal comparison. Deep Sea Res. Part II Top. Stud. Oceanogr. 59–60, 78–92. <https://doi.org/10.1016/j.dsrt.2011.07.004>.
- Waters, K.J., Smith, R.C., 1992. Palmer LTER: a sampling grid for the Palmer LTER program. Antarct. J. U. S. 27, 236–239.

- Wiebe, P.H., Morton, A.W., Bradley, A.M., Backus, R.H., Craddock, J.E., Barber, V., Cowles, T.J., Flierl, G.R., 1985. New development in the MOCNESS, an apparatus for sampling zooplankton and micronekton. *Mar. Biol.* 87, 313–323. <https://doi.org/10.1007/BF00397811>.
- Williams, R., Robins, D.B., 1981. Seasonal variability in abundance and vertical distribution of *Parathemisto gaudichaudi* (Amphipoda: Hyperiidea) in the North East Atlantic Ocean. *Mar. Ecol. Prog. Ser.* 4, 289–298.
- Wilson, S.E., Steinberg, D.K., Buesseler, K.O., 2008. Changes in fecal pellet characteristics with depth as indicators of zooplankton repackaging of particles in the mesopelagic zone of the subtropical and subarctic North Pacific Ocean. *Deep Sea Res. Part II Top. Stud. Oceanogr.* 55, 1636–1647. <https://doi.org/10.1016/j.dsr2.2008.04.019>.
- Zaret, T.M., Suffern, J.S., 1976. Vertical migration in zooplankton as a predator avoidance mechanism. *Limnol. Oceanogr.* 21, 804–813. <https://doi.org/10.4319/lo.1976.21.6.0804>.
- Zhou, M., Dorland, R.D., 2004. Aggregation and vertical migration behavior of *Euphausia superba*. *Deep Sea Res. Part II Top. Stud. Oceanogr.* 51, 2119–2137. <https://doi.org/10.1016/j.dsr2.2004.07.009>.